

PREFACE

Dear friends!

We are happy to present you the Booklet of Preparatory problems. This year its structure was changed according to the recommendations of the International Steering Committee (ISC). Besides problems and worked solutions, you will find in the Booklet:

- Minutes of the ISC Meeting held in Moscow on December 7-10, 2006
- Proposed Agenda of the Business part of the 1st Jury Meeting
- The Syllabi for the practical and theoretical parts
- The Safety rules and recommendations set by the International Jury
- The hazard warning symbols, their designations and explanations, R-ratings and S-provisions

All these documents can be also found at our official website www.icho39.chem.msu.ru

In the Booklet you will also find the list of level 3 areas organizers would like students to be acquainted with. Note that this is not a simple enumeration of level 3 topics from the Syllabus. It is rather an informal presentation of fields of advanced difficulty that will be addressed at the forthcoming Olympiad. We publish the list to make your preparation more effective.

Note to mentors:

Please study the Minutes and Proposed Agenda carefully, since many important questions have been discussed at the ISC Meeting in December 2006, and the debates will be continued in Moscow.

We place a great importance on safety. In the section preceding the practical preparatory problems you will find safety precautions and procedures to be followed. At the registration in Moscow we will ask the Head mentors to sign a form stating that their students are aware of safety rules and adequately trained to follow them. Prior to the practical exam all students will have to read and sign safety instructions translated in their languages of choice.

Despite our great proof reading efforts, some mistakes and misprints are still possible. We appreciate your understanding and will be happy to get feedback. Please address your comments to secretary@icho39.chem.msu.ru

Note to students:

Members of the Science Committee really did their best to prepare interesting tasks. The set covers all major parts of modern chemistry, though most of tasks can be solved by applying a basic knowledge of chemistry. Answers to the questions that will be posted at the website by the end of May are very detailed to give you an opportunity to learn the backgrounds. Elaborating the tasks, we intended not only to announce particular chemistry fields, but also to make the material challenging and give you an idea of the structure and spirit of problems which you will see at the competition in July. We were also keen to provide you with sufficient material for training. Enjoy solving the tasks and please do not forget that **CHEMISTRY IS ART, SCIENCE, and FUN!**

Welcome to the International Chemistry Olympiad in Moscow!

Members of IChO-2007 Science Committee

LIST OF FIELDS OF ADVANCED DIFFICULTY

| Problem(s) | Field | Subfields |
|------------|-------------------------------------|---|
| 1 | Periodic trends | – |
| 2, 3 | Chemical bonding, quantum mechanics | Superposition principle. Molecular orbitals. Periodic wave functions. Uncertainty principle. |
| 4 | Photochemistry | Energy diagram of a chemical reaction. Activation energy. Relationship between energy and wavelength of light. |
| | Quantum mechanics | Particle-in-a-box model. |
| 5-7 | Equilibrium | Surface tension. Gibbs energy and its dependence on pressure for pure substance. The temperature dependence of the saturated vapor pressure. Relationship between $\Delta_r G^\ominus$ and equilibrium constant K . Using ΔG to predict direction of natural change. Dependence of $\Delta_r G$ on partial pressures of reactants and products. Le Chatelier's principle. |
| 8 | Phase diagrams, equations of state | Single component phase diagrams. Critical point. Van der Waals gas law. |
| 9-11 | Chemical kinetics | Determination of the reaction order. Rate-determining step. Steady-state approximation. Calculation of activation energy. Kinetic equations and kinetic curves. Autocatalysis. Enantiomeric enrichment. First-order reactions: Dependence of concentration on time, half-life. Carbon dating. |
| | Carbonyl compounds | Addition reactions. Stereochemistry: enantiomers. |
| 12-14 | Inorganic chemistry of elements | Fe(II) and Fe(III), redox processes, cyanide and tartrate complexes, hydroxides. MnO_4^- as an oxidizing agent in acidic media. As(III) and As(V), redox processes. Compounds of sulfur in lower oxidation states, oxidation with iodine. Zinc, sulfide and carbonate, their solubility. Phosphates, their thermal decomposition. |
| | Electrochemistry | Standard electrode potentials. Nernst equation. EMF. Direction of redox processes. |
| | Chemical equilibria | Acid-base and precipitation equilibria, calculation of pH, K_{sp} in complex mixtures. |
| | Analytical chemistry | Redox titration (direct and back-titration). Stoichiometric calculations. |
| | Carbonyl compounds | Nucleophilic addition of HSO_3^- . |

| | | |
|-------|---|---|
| 15-17 | Chemical bonding | VSEPR-concept (factors affecting distortion of an ideal polyhedron). Crystal Field Theory of coordination compounds. Calculation of Crystal Field Stabilization Energy. |
| | Solid state chemistry | Unit cell. Coordination number. Miller indices. Bragg's Law. Types of close packings. Calculation of density of packings. X-ray diffraction for f.c.c. lattice. NaCl, spinel, and perovskite structure. |
| | Equilibrium | Hard and Soft Acids and Bases (HSAB) concept. Hydrolysis, calculation of pH. Osmotic pressure. Free energy definition. Relationship between ΔG° and equilibrium constant K . Using ΔG to predict direction of natural change. |
| | Inorganic chemistry of elements | Group 14: oxocompounds ((+4) oxidation state of the elements). Group 15: oxoacids with the element having (+1), (+3) or (+5) oxidation states; structure of the acids; pK_a trends. Polymerization of oxoacids (oxoanions). Transition metals: tetrahedral and octahedral complexes of Co and Cr. |
| 18-20 | Carbonyl compounds | Aldehydes, ketones, carboxylic acid derivatives: properties, keto-enol tautomerism, enolates and enol derivatives. |
| | Condensations of carbonyl compounds | General principles, mechanism of base-catalyzed condensations. |
| | Concerted pericyclic reactions | General principles and common types of pericyclic processes. |
| 21-24 | Amino acids and peptides (without proteins) | Structure, sequencing, chemical properties of carboxyl, amino and functional side groups. |
| | Lipids | Structure, physical and chemical properties, synthesis and degradation. |
| | Bases, nucleosides and nucleotides: (without nucleic acids) | Structure and properties. |
| | Enzymes | Nomenclature, mechanisms of catalysis, specificity. |
| | Physico-chemical methods | ^1H NMR and mass spectrometry. |
| 25-27 | Polymerization | Mechanisms, stages, kinetics, characteristics of obtained polymers |
| | Monomer structure and reactivity in polymerization | Inductive and mesomeric effects, ring strain, solvent effect, etc. |
| | Copolymers | Synthesis, architecture, distribution of units, properties. |
| | ^1H NMR for studying polymers | Common ranges of chemical shifts of typical functional groups and simple fragments, integration of signals. |
| 28 | Quantum mechanics | Energy diagram of a chemical reaction. Tunneling. Relationship between frequency, energy and wavelength of light. |

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The 39th International Chemistry Olympiad

Chemistry: art, science and fun



PREPARATORY PROBLEMS (Theoretical)

July 15-24, 2007

Moscow, Russia

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Problem 1. ON THE BORDERS OF THE PERIODIC SYSTEM

| | | | | | |
|------|--------|----------|---------|----------|----------|
| | | | Ti=50 | Zr=90 | ?=180. |
| | | | V=51 | Nb=94 | Ta=182. |
| | | | Cr=52 | Mo=96 | W=186. |
| | | | Mn=55 | Rh=104,4 | Pt=197,4 |
| | | | Fe=56 | Ru=104,4 | Ir=198. |
| | | Ni=Co=59 | | Pt=106,4 | Os=199. |
| | | Cu=63,4 | | Ag=108 | Hg=200. |
| H=1 | Be=9,4 | Mg=24 | Zn=65,3 | Cd=112 | |
| | B=11 | Al=27,4 | ?=68 | Ur=116 | Au=197? |
| | C=12 | Si=28 | ?=70 | Sn=118 | |
| | N=14 | P=31 | As=75 | Sb=122 | Bi=210 |
| | O=16 | S=32 | Se=79,4 | Te=128? | |
| | F=19 | Cl=35,4 | Br=80 | I=127 | |
| Li=7 | Na=23 | K=39 | Rb=85,4 | Cs=133 | Tl=204 |
| | | Ca=40 | Sr=87,4 | Ba=137 | Pb=207. |
| | | ?=45 | Ce=92 | | |
| | | ?Er=56 | La=94 | | |
| | | ?Yt=60 | Di=95 | | |
| | | ?In=75,4 | Th=118? | | |

The first Periodic system of the elements was proposed in 1869 by the Russian chemist D.I. Mendeleev, who arranged all the known chemical elements in the order of increasing atomic mass. In 1871 Mendeleev published the article «The natural system of the elements and its application to the prediction of properties of yet undiscovered elements» in the «Journal of the Russian Chemical Society». In that article Mendeleev described in detail the properties of three unknown elements that were ekaboron (Eb), ekaaluminum (Ea), and ekasilicon (Es). All of them were discovered in the next 15 years.

1. What are the present names of the three elements predicted by Mendeleev? Interestingly, all three names have a geographical origin.

The first Periodic system listed 66 elements only, of which three were unknown. In the present-day system there are 118 elements. The last, 118th element was discovered in 2005 during the collaborative studies by the Joint Institute for Nuclear Research (Russia) and the Livermore National Laboratory (USA). After the collisions of calcium-48 nuclei with the target containing californium-249 nuclei three cascades of α -decay were detected, that started from the 118th element with the mass number 294.

2. Write the balanced equations of the nuclear reactions of: i) the synthesis and ii) the α -decay of the 118th element.

3. To which group of the Periodic system does the 118th element belong? Give its electron configuration using a noble gas with the *spdf* notation.

4. Based on the properties of the same-group analogs of the 118th element and using extrapolation predict the following properties of the 118th element: i) melting point; ii) boiling point, iii) atomic radius, iv) first ionization energy, v) the formula of the oxide of the 118th element in its highest oxidation state.

Problem 2. SCHRÖDINGER CAT AND CHEMISTRY

Many chemical phenomena can be explained by physical theories. The main theory for chemistry is quantum mechanics, which gives the solid foundation for the observed chemical periodicity. One of the cornerstones of quantum mechanics is the superposition principle that says:

“If a quantum system can be found in the states 1 and 2 described by wavefunctions Ψ_1 and Ψ_2 , it can also be found in a mixed state with the wavefunction

$$\Psi = c_1\Psi_1 + c_2\Psi_2,$$

where factors c_1 and c_2 characterize the contributions of the pure states 1 and 2 to the mixed state”.

The sum or difference of some wave functions taken with certain factors is called a superposition (a linear combination) of these functions.

In a mixed state the quantum system exists in both pure states simultaneously. When you perform some measurement on the system being in the mixed state, this measurement transfers the system to one of the pure states. We can never predict the specific final state; it is determined by the probability laws. The probability of any of the final states after measurement is proportional to the square of the modulus of the corresponding factor:

$$p_1 \sim |c_1|^2, \quad p_2 \sim |c_2|^2.$$

Of course, the probability to find the system in either of the states is unity:

$$p_1 + p_2 = 1.$$

The superposition principle is applicable to quantum systems only and is not valid when applied to macrosystems. To illustrate this idea, E. Schrödinger proposed the following mental experiment. Consider the Geiger counter which detects the entering electrons. The counter is connected to a device which breaks the glass with the poison when the particle enters the counter. Near the glass is a live cat. If the particle enters the counter, the cat is poisoned. But if the counter did not perform the measurement and is in the mixed state between the detected and undetected particle then the state of the cat is a superposition of life and death. Evidently, this is nonsense: the cat can be either alive or dead.

In chemistry, the superposition principle is used in the theories of hybridization, resonance, and molecular orbitals.

The superposition principle in theory of hybridization.

1. An sp^3 -hybrid atomic orbital is a linear combination of one s and three p-orbitals:

$$\Psi_{sp^3} = c_1\Psi_s + c_2\Psi_{p_x} + c_3\Psi_{p_y} + c_4\Psi_{p_z}.$$

i) If we assume that all the orbitals make an equal contribution to a hybrid orbital, what are the absolute values of the coefficients $c_1 - c_4$?

ii) Similarly, find the absolute values of the coefficients $c_1 - c_3$ for an sp^2 hybrid orbital.

The superposition principle in molecular orbital theory.

2. The molecular orbital for the ground state of H_2^+ molecule ion has the form:

$$\Psi = \frac{1}{\sqrt{2}}\Psi_{1s}^a + \frac{1}{\sqrt{2}}\Psi_{1s}^b,$$

where a and b denote hydrogen atoms. What is the probability to find an electron on the 1s-orbital of the atom a ?

The superposition principle in theory of resonance.

3. Covalent bonds have a partial ionic character. Thus the wavefunction of a hydrogen halide bond can be presented as a linear combination of two wavefunctions characterizing its ionic ($\Psi_{H^+Hal^-}$) and covalent ($\Psi_{H:Hal}$) states:

$$\Psi_{HHal} = c_{cov}\Psi_{H:Hal} + c_{ion}\Psi_{H^+Hal^-}$$

L. Pauling in his famous book «The nature of the chemical bond» (1947) claimed that in the HCl molecule the chemical bond is 17% ionic in character. Find the absolute values of c_{cov} and c_{ion} for HCl.

4. One of the benzene wavefunctions can be presented as a linear combination of wavefunctions that correspond to two Kekule and three Dewar structures:

$$\Psi_{\text{C}_6\text{H}_6} = \frac{\sqrt{2}}{\sqrt{5}} \Psi_{\text{Kekule}} + \frac{\sqrt{2}}{\sqrt{5}} \Psi_{\text{Kekule}} + \frac{1}{\sqrt{15}} \Psi_{\text{Dewar}} + \frac{1}{\sqrt{15}} \Psi_{\text{Dewar}} + \frac{1}{\sqrt{15}} \Psi_{\text{Dewar}}$$

What is the total contribution of the Kekule structures to this electronic state of benzene?

In chemical reactions molecular structure changes over time so that the electronic state of a molecule is a function of time. In some cases structure of a molecule can be presented by a superposition of the initial and final states with time-dependent coefficients.

Let's assume that a molecule oscillates between two pure states, one with a wave function Ψ_1 , and another with a wavefunction Ψ_2 , with the frequency ω . Initially ($t = 0$) the molecule is in the pure first state and after a half-period ($t = \pi/\omega$) – in the second pure state.

5. Find the time-dependent coefficients of the superposition of these states describing the electronic structure of the molecule. Write the total wave function at a quarter of a period.

Problem 3. QUANTUM UNCERTAINTY

One of the main quantum laws relates the uncertainties of position Δx and momentum Δp of quantum particles. The uncertainty product cannot be less than a fixed value – a half of Planck's constant:

$$\Delta x \cdot \Delta p \geq \frac{\hbar}{2}$$

where momentum is the product of mass and velocity: $p = mV$, the Planck's constant is $\hbar = 1.05 \cdot 10^{-34}$ J·s.

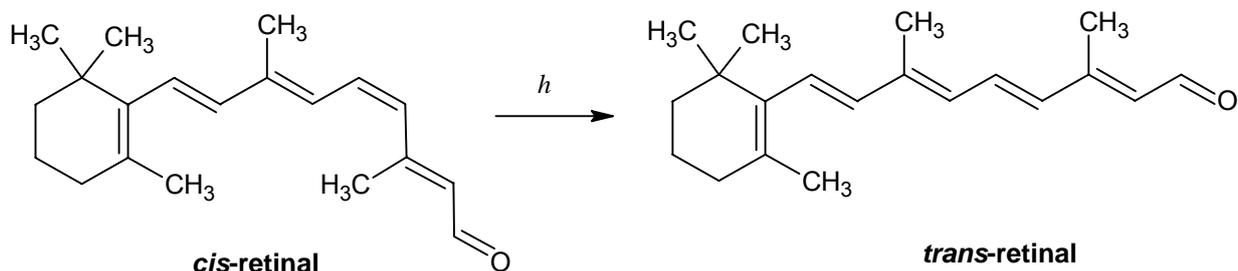
1. Without performing calculations arrange the following particles in the order of increasing minimal uncertainty of velocity, ΔV_{\min} :

- an electron in a H_2 molecule;
- a H atom in a H_2 molecule;
- a proton in the carbon nucleus;
- a H_2 molecule within a nanotube;
- a O_2 molecule in the room of 5 m width.

2. For the first and the last particles from the list above calculate ΔV_{\min} . Take the necessary reference data from handbooks or Internet.

Problem 4. QUANTUM CHEMISTRY OF VISION

The first step in the very complex mechanism of vision is the photoinduced *cis* \rightarrow *trans* isomerization of the chromophore retinal embedded in rhodopsin molecules. Absorption of visible light by *cis*-retinal causes a change of the configuration of a double bond:



1. Show the double bond, which participates in the *cis-trans*-isomerization. Indicate the reaction coordinate.

2. Energies of the reactant and the product were found to be periodic functions of the reaction coordinate x :

$$E_{\text{cis}}(x) = 1.79 \cdot (1 - \cos(x)),$$

$$E_{\text{trans}}(x) = 1.94 + 0.54 \cdot \cos(x).$$

Energies are in eV ($1 \text{ eV} = 1.60 \cdot 10^{-19} \text{ J} = 96500 \text{ J/mol}$), $x = 0$ corresponds to the reactant, $x = \pi$ – to the product. Draw the energy diagram for this reaction. Determine the energy change for the reaction and its activation energy in kJ/mol.

3. What is the largest wavelength of light that can be absorbed by *cis*-retinal?

Let us apply the “particle-in-a-box” model to the electrons present in the conjugated system of *cis*-retinal. Energy levels of a particle of the mass m locked in an one-dimensional box of the width l are given by:

$$E_n = \frac{h^2 n^2}{8ml^2}, \quad n = 1, 2, \dots$$

4. What is the number of electrons in the conjugated system of *cis*-retinal?

5. Based on your answers on questions (3)-(4) and using the formula above calculate l . How does this value compare with the structure of retinal molecule?

Problem 5. NANOPARTICLES AND NANOPHASES

Nanochemistry has sparked much excitement in the recent years and a large amount of research has been dedicated to understanding of nanomaterials. Single-walled carbon nanotubes (SWNTs) are a universally known example of such materials. SWNT can be thought of as a sheet of graphite rolled into a seamless cylinder ($d \approx 1.5 \text{ nm}$). These cylindrical carbon “molecules” might provide components for molecular electronic devices of the future.

The properties of nanometer-scale materials are size- and shape-dependent.

Saturated vapor pressure of a small spherical particle (crystalline or liquid) is higher than that of the bulk phase of the same material. At equilibrium the molar Gibbs functions (G)

of the condensed phase (G_{bulk}) and vapor (G_{vap}) are equal. Equation (1) determines the saturated vapor pressure, p , above a bulk phase

$$G_{\text{bulk}} = G_{\text{vap}} = G^{\circ}_{\text{vap}} + RT \ln p, \quad (1)$$

G°_{vap} is the standard molar Gibbs energy of vapor at standard pressure $p = 1$ bar.

The substance inside a small spherical sample is under excess pressure, caused by surface tension:

$$\Delta P_{\text{in}} = 2\sigma / r$$

r – the radius of the spherical sample, σ – the surface tension at the “condensed phase-vapor” interface. The increase of the internal pressure results in a change in the molar Gibbs energy of the substance inside the spherical sample. This molar Gibbs energy G^*_{sph} is larger than G_{bulk} . The difference in the Gibbs energy of the spherical sample and the bulk phase is equal to $\Delta P_{\text{in}} V$:

$$G^*_{\text{sph}} = G_{\text{bulk}} + \Delta P_{\text{in}} V = G_{\text{bulk}} + 2\sigma V / r, \quad (2)$$

V is the molar volume of the liquid or solid substance. Therefore from equation (1)

$$G^*_{\text{sph}} = G_{\text{bulk}} + 2\sigma V / r = G_{\text{vap}} = G^{\circ}_{\text{vap}} + RT \ln p^* \quad (3)$$

p^* is the saturated vapor pressure of the spherical sample with the radius r .

1. The saturated vapor pressure of water at $T = 298$ K is $3.15 \cdot 10^{-2}$ bar. Calculate the saturated vapor pressure of the spherical droplets of water with the radius of: i) $1 \mu\text{m}$ and ii) 1 nm . The surface tension at the liquid-vapor interface of water is 0.072 J/m^2 .

Assuming that the substance retains properties of a bulk while the difference between its saturated vapor pressure and the saturated pressure of the bulk is less than 1%, what is the minimum radius of the spherical sample that can still be considered as a bulk phase? How many molecules of water are there in such a droplet?

2. Few droplets of mercury were put inside a SWNT maintained at 400 K. What is the minimum vapor pressure of mercury inside the tube? The saturated vapor pressure of bulk mercury is $1.38 \cdot 10^{-3}$ bar, the density of mercury $\rho(\text{Hg}) = 13.5 \text{ g/cm}^3$, the surface tension at the liquid-vapor interface of mercury is 0.484 J/m^2 at the given temperature.

3. The boiling point of benzene at the standard atmospheric pressure is $T_b = 353.3$ K. The temperature dependence of the saturated vapor pressure of benzene near the boiling point is given by the equation

$$\ln p(T) = -\frac{\Delta H_{\text{vap}}}{RT} + \text{const} \quad (4)$$

where $\Delta H_{\text{vap}} = 30720$ J/mol is the enthalpy of vaporization of benzene. Estimate the boiling point (T^*) of the finely dispersed liquid benzene at the standard atmospheric pressure if the sample consists of droplets with the radius $r = 50$ nm. The surface tension of benzene near the boiling point is 0.021 J/m² and its density is 0.814 g/cm³.

4. In general, properties of the bulk and nano-sized material composed by one and the same substance A are different. Which of the following thermodynamic constants will decrease when passing from the bulk to the nano-scaled material?

- 1) Solubility of A in any solvent;
- 2) the boiling temperature at atmospheric pressure;
- 3) the saturated vapor pressure over solid substance A;
- 4) the equilibrium constant of a chemical reaction, where A is a reagent;
- 5) the equilibrium constant of a chemical reaction, where A is a product.

Problem 6. IN WHICH DIRECTION DOES A CHEMICAL REACTION PROCEED?

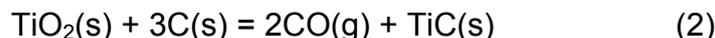
The natural tendency of any chemical reaction to proceed in a certain direction at constant temperature and pressure is determined by the sign of the Gibbs energy of the reaction, ΔG . This is the universal principle. If $\Delta G < 0$, the reaction can proceed predominantly in the forward direction (a product-favored reaction). If $\Delta G > 0$ the reaction can proceed predominantly in the reverse direction (a reactant-favored reaction). When $\Delta G = 0$ the reaction is at equilibrium.

The standard reaction Gibbs energy, ΔG° , can be calculated from the tabulated Gibbs energies of formation of the reactants and products (see the Table).

1. Calculate the equilibrium constant of reaction (1) at 1627 °C. Can the reaction proceed predominantly in the forward direction if the initial partial pressure of O₂ is below 1.00 Torr?



2. The standard Gibbs energy of the reaction



is positive at 727 °C. Calculate the equilibrium pressure of CO at 727 °C. What should be the reaction conditions to allow for the forward reaction to be the predominant process at this temperature if this is possible at all?

3. Calculate the standard Gibbs energy of the reaction



at 300 K. Can the forward reaction be the predominant process under the following conditions: $p(\text{NH}_3) = 1.0 \text{ atm}$, $p(\text{H}_2) = 0.50 \text{ atm}$, $p(\text{N}_2) = 3.0 \text{ atm}$?

In fact the reaction does not occur at 300 K at a noticeable rate. Why?

Table 1. Gibbs energies of formation*.

| Substance | $t, \text{ }^\circ\text{C}$ | $\Delta_f G^\circ, \text{ kJ/mol}$ |
|------------------|-----------------------------|------------------------------------|
| NiO | 1627 | -72.1 |
| TiO ₂ | 727 | -757.8 |
| TiC | 727 | -162.6 |
| CO | 727 | -200.2 |
| NH ₃ | 27 | -16.26 |

*The standard pressure – 1atm, JANAF Tables.

Problem 7. LE CHATELIER'S PRINCIPLE

Le Chatelier's principle states that

«Every system in the state of equilibrium when subjected to a perturbation responds in a way that tends to eliminate the effect» (P.W. Atkins "Physical Chemistry").

Let us see how this principle works. Let a chemical equilibrium be established in the following reaction between the ideal gases:



At the temperature of $T = 400$ K partial pressures of reactants and product are respectively: $p(\text{H}_2) = 0.376$ bar, $p(\text{N}_2) = 0.125$ bar, $p(\text{NH}_3) = 0.499$ bar.

The equilibrium was disturbed. Let this disturbance be

- a) increase of the total pressure in the system at constant temperature,
- b) increase of the amount of NH_3 in the system at constant total pressure and temperature,
- c) small increase of the amount of N_2 in the system at constant total pressure and temperature,
- d) small increase of the amount of H_2 in the system at constant total pressure and temperature.

1. Calculate the standard Gibbs energy for the reaction (1) at $T = 400$ K.
2. Write down the expression for the Gibbs energy of reaction (1) for any pressure of reactants and product after perturbation. This expression is called the isotherm of chemical reaction.
3. Using the equation of isotherm from question 2 determine in which direction the reaction (1) will predominantly proceed after the disturbance of equilibrium as indicated in (a)-(d).
4. Will the answers to question 3 change, if the initial equilibrium partial pressures in the system are: $p(\text{H}_2) = 0.111$ bar, $p(\text{N}_2) = 0.700$ bar, $p(\text{NH}_3) = 0.189$ bar? Assume that temperature and total pressure in the system are the same as in questions 1–3.

Problem 8. DMITRY IVANOVICH MENDELEEV: WHAT BESIDES THE PERIODIC TABLE?

The Russian chemist D. Mendeleev is known for his Periodic Table of elements. This discovery made him famous worldwide. Dmitry Mendeleev has carried out some other interesting studies as well. Consider two of them.



1. Mendeleev was the first to state that every substance has “the temperature of the absolute boiling”. Above this temperature “the substance will stay in the gas phase no matter how high the pressure is”. According to Mendeleev “the temperature of the absolute boiling of water” is 543 °C.

- a) What is “the temperature of the absolute boiling”?
- b) Indicate the temperature of the absolute boiling in the P - T phase diagram of water.
- c) Calculate the temperature of the absolute boiling of water from the Van der Waals equation of state:

$$\left(p + \frac{a}{V^2}\right)(V - b) = RT,$$

For H_2O , $a = 5.464 \text{ l}^2 \cdot \text{atm} \cdot \text{mol}^{-2}$, $b = 0.03049 \text{ l} \cdot \text{mol}^{-1}$.

2. In Russia many people believe that D. Mendeleev invented the recipe of the famous drink “Russian vodka”. We have a chance to check this legend.

The fact is that in his Ph.D. thesis Mendeleev characterized some properties of the binary system “ethanol-water”. He measured the density ρ of a series of binary solutions of various compositions W , where $W(\%)$ is the weight percent of ethanol in the mixture. The derivative $d\rho / dW$ is presented in Fig.1 as a function of W .

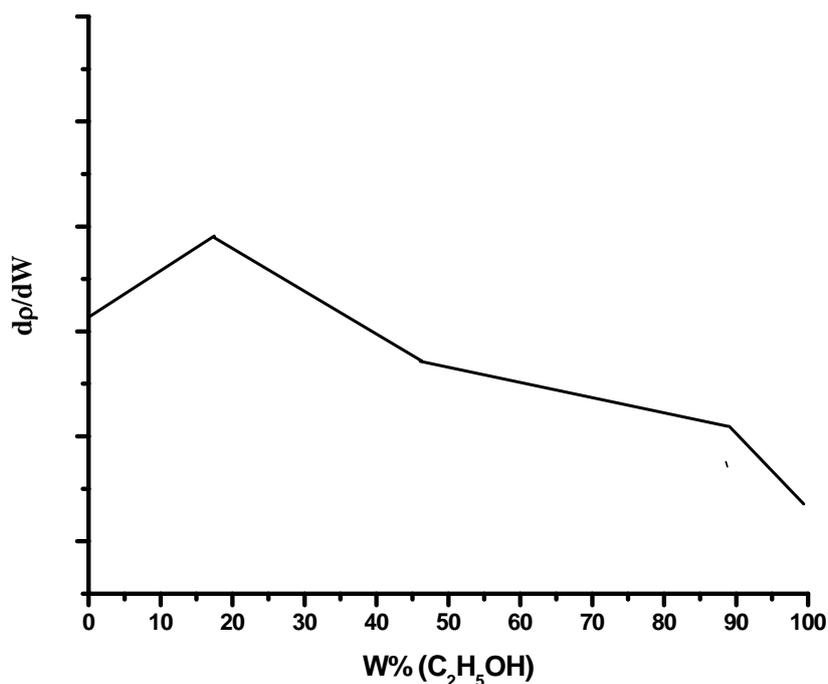


Fig. 1. Experimental results obtained by Mendeleev

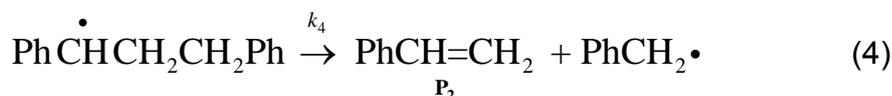
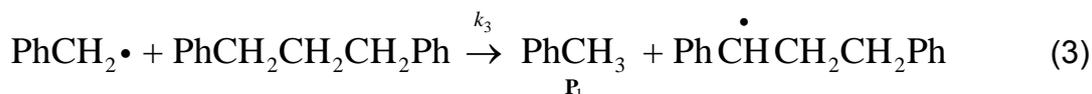
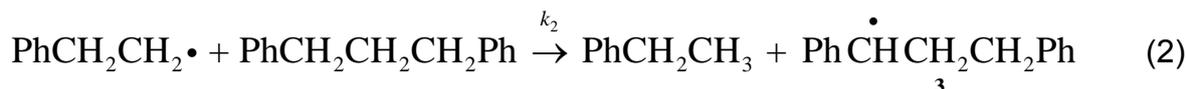
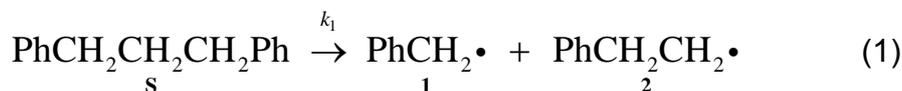
The curve markedly changes the slope three times. According to D. Mendeleev these three special points correspond to the compositions of the weakly bonded chemical compounds, “hydrates of ethanol”.

- a) What are the chemical formulas of “the hydrates of the ethanol”?
- b) Does the composition of any of the “hydrates” resemble the recipe of vodka (40 volume percent of C_2H_5OH)? The density of ethanol is $0.794 \text{ g}\cdot\text{cm}^{-3}$. Decide whether or not Dmitry Mendeleev took part in “the discovery of Russian vodka”.

Problem 9. KINETICS OF A FREE RADICAL REACTION

Pyrolysis is an important industrial process for conversion of coal to liquid fuels and chemical feedstocks. The structure of coal can be viewed as a three-dimensional network of polycyclic aromatic building blocks joined together by short aliphatic bridges. In model pyrolysis studies, α,ω -diphenylalkanes are sometimes used as model compounds for coal.

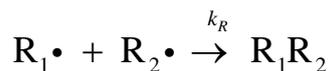
Thermal decomposition of 1,3-diphenylpropane gives toluene and styrene as the major products and ethylbenzene and other hydrocarbons as byproducts. The following mechanism of decomposition has been proposed (the first step is the slowest):



1. Applying the steady-state approximation for the radical **2**, derive the rate equation for the side reaction of ethylbenzene formation.

2. What is the ratio between the steady-state concentrations of the radicals **1** and **3**?

Additionally, two free radicals can recombine. The rate constant of recombination k_R is supposed to be the same for all radicals.



3. Why could we neglect these reactions in the steady-state equations in questions 1 and 2?

4. One of the radicals is present in the reaction mixture at much higher concentration than others. This radical is:

- $\text{Ph}\dot{\text{C}}\text{HCH}_2\text{CH}_2\text{Ph}$, because it is the most stable one;
- $\text{PhCH}_2\cdot$, because the rate constant of β -scission reaction (4) is higher than the rate constant of chain propagation reaction (3);
- $\text{PhCH}_2\text{CH}_2\cdot$, because it accumulates in the system.

5. Obtain the rate equation for toluene formation. Determine the reaction order. Express the effective activation energy via the activation energies of elementary steps.

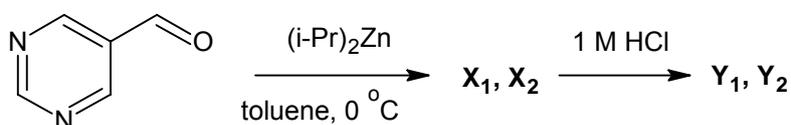
Problem 10. ASYMMETRIC AUTOCATALYSIS – AMPLIFICATION OF CHIRAL ASYMMETRY

Living nature is homochiral: almost all natural amino acids have L-configuration, sugars – D-configuration. One of the possible explanations of this phenomenon is based on the concept of asymmetric autocatalysis. In some reactions chiral products can serve as catalysts of their own formation: the larger is the content of one of the enantiomers the faster is its synthesis.

1. The simplest equation for autocatalysis is: $A + P \rightarrow 2P$, where P is product. Reaction can be performed under various conditions: either in a closed system when reagents are mixed only once, or in an open system where reagent A is being continuously added to the mixture so that its concentration is maintained constant.

Write the kinetic equations and draw the kinetic curves for product P in the closed and open systems. Assume that the initial concentration of P is non-zero but small.

The first reaction of asymmetric autocatalysis was discovered in the early 1990-s. Addition of diisopropylzinc to pyrimidine-5-carbaldehyde in toluene leads to the mixture of enantiomers X_1 and X_2 , which after hydrolysis is transformed to enantiomeric alcohols Y_1 and Y_2 :



2. Draw the structure of enantiomeric pairs **X** and **Y**, and show the configuration of the stereocenter.

It turned out that the presence of small amounts of any product (**Y**₁ or **Y**₂) selectively accelerates the formation of that specific product which leads to enantiomeric enrichment of the reaction mixture. Suppose that the yield of each product is proportional to the square of its molar fraction in the mixture of alcohols prior to synthesis.

3. To 1 mmol of mixture **Y**₁ and **Y**₂, containing 55% of **Y**₁, 1 mmol of aldehyde and 1 mmol of diisopropylzinc are added several times. Assuming that total reaction yield is 100%, calculate how many times we should add the reagents to enrich the mixture of alcohols up to: a) 70%, b) 90%, c) 99% of **Y**₁.

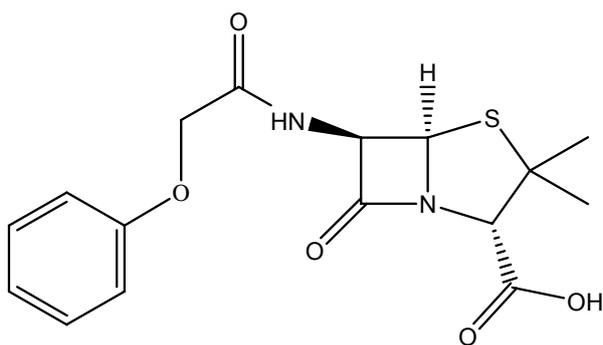
Note. You need to write a small iteration program.

Problem 11. RADIOCARBON DATING

The carbon-14, a radioactive isotope of carbon, is often used to date archaeological, geological, and hydrogeological samples. The half-life of ¹⁴C is $t_{1/2} = 5730$ years, but in calculations of the age of samples, a different value of half-life, $t'_{1/2} = 5568$ years, is used. The ¹⁴C is produced from nitrogen in the atmosphere under the action of cosmic rays. It can be included in the organisms of plants and animals through the photosynthesis and the food chains. The radiocarbon content in living organisms is nearly constant with the activity of ¹⁴C being 230 Bq per kg of carbon. After death of an organism, the carbon exchange stops and the ¹⁴C content starts decreasing continually.

1. Give the balanced reaction equations of formation and decay of ¹⁴C.
2. Activity of radiocarbon in a sample of cloth from an Egyptian pyramid corresponds to 480 disintegrations per hour per gram of carbon. What is the age of the cloth?

In another pyramid, a white powder was found. Analysis showed it was a pure phenoxymethylpenicillin (Penicillin V):



Commercial phenoxymethylpenicillin is produced by microorganisms cultured in a medium containing carbohydrates (lactose, glucose, sucrose), cornsteep liquor, mineral salts and phenoxyacetic acid.

It was decided to determine the radiocarbon content to estimate the age of the powder. The $^{14}\text{C}/^{12}\text{C}$ ratio determined from mass-spectrometry measurements amounts to $6.0 \cdot 10^{-13}$.

3. The archaeologists estimated the age of the powder from the radioactive decay law. What was the production date they obtained?
4. Explain this result. When was the powder produced in reality?

Constants were taken from:

Lloyd A. Currie. The Remarkable Metrological History of Radiocarbon Dating. // J. Res. Natl. Inst. Stand. Technol. 109, 185-217 (2004)

Problem 12. IRON DETERMINATION

Iron is one of the most important elements necessary for the support of the vital functions of human organism. Its deficiency may cause anemia for treatment of which Fe(II)

supplementation is usually employed. The therapeutic effect of Fe(III) compounds is much less pronounced.

Fe(II) is a fairly strong reducing agent which can be readily oxidized to Fe(III). Therefore methods for separate determination of Fe(II) and Fe(III) as well as for the determination of the total iron content are needed for quality control of pharmaceuticals. Here we will see how this problem can be solved.

1. Prior to determination of the total iron content it is usually transformed quantitatively either to Fe(II) or to Fe(III). Using standard redox potentials given below establish which of the oxidizing agents listed can oxidize Fe(II) to Fe(III) under standard conditions. Write down the balanced net ionic equations of corresponding reactions.

| oxidized form | reduced form | E° , V |
|--------------------------------------|-----------------------------------|---------------|
| Fe^{3+} | Fe^{2+} | +0.77 |
| HNO_3 | $\text{NO} (+\text{H}_2\text{O})$ | +0.96 |
| $\text{H}_2\text{O}_2 (+\text{H}^+)$ | H_2O | +1.77 |
| I_2 | I^- | +0.54 |
| Br_2 | Br^- | +1.09 |

2. After oxidation of all the iron to Fe(III) its total amount can be determined by precipitation of iron in the form of $\text{Fe}(\text{OH})_3$ followed by annealing of the precipitate to Fe_2O_3 and weighing.

- Estimate the pH of 0.010 M FeCl_3 in water. Assume that $\text{Fe}(\text{OH})_2^{3+}$ cation is a monoprotic acid with the dissociation constant $K_a = 6.3 \cdot 10^{-3}$.
- Calculate the pH necessary to begin precipitation of $\text{Fe}(\text{OH})_3$ from the solution above. Solubility product of $\text{Fe}(\text{OH})_3$ is $K_{sp} = 6.3 \cdot 10^{-38}$.
- At what pH value precipitation of $\text{Fe}(\text{OH})_3$ from 100.0 mL of 0.010 M FeCl_3 will be complete? Consider the precipitation as complete if no more than 0.2 mg Fe remains in solution.

Note. All the pH values should be estimated with accuracy of 0.1 units pH. Neglect the effect of ionic strength.

3. Fe(II) can be determined in the presence of Fe(III) by titration with KMnO_4 solution in acidic media. Since aqueous solutions of KMnO_4 tends to decompose slowly over time, the exact concentration of KMnO_4 has to be found immediately before determination of Fe(II). This is usually done by titration with KMnO_4 of a solution of a primary standard, a pure substance of known composition. Such standard solution can be prepared by dissolving an exact amount of the primary standard in water in a volumetric flask of an exactly known volume.

For the titration of 10.00 mL of a primary standard solution containing 0.2483 g of As_2O_3 in 100.0 mL of water 12.79 mL of KMnO_4 solution were used, whereas for titration of 15.00 mL of the solution containing 2.505 g Fe per liter were used 11.80 mL of that same solution of KMnO_4 . What fraction of iron in the sample was present in the form of Fe(II)?

4. To a solution containing Fe(II) and Fe(III) tartaric acid was added. The solution was neutralized with aqueous ammonia and then excess KCN was added. The potential of the platinum electrode immersed in that solution was found to be +0.132 V against saturated calomel electrode.

a) Assuming that all iron in the last solution was present in the form of $\text{Fe}(\text{CN})_6^{7-}$, calculate the fraction of iron present in the form of Fe(II) in the original sample. Standard redox potential of $\text{Fe}(\text{CN})_6^{3-}/\text{Fe}(\text{CN})_6^{4-}$ is +0.364 V. Potential of saturated calomel electrode is +0.241 V. The temperature of the sample solution is 25 °C.

b) What concurrent reactions were prevented by the addition of tartaric acid and ammonia to the sample solution? Write down the net ionic equations of those reactions.

Problem 13. SULFUR DETERMINATION

Compounds of sulfur in its lower oxidation states are present in many industrial wastes (metallurgy, production of paper, chemical) and are dangerous ecotoxicants. The prevalent forms of sulfur in lower oxidation states in solutions are S^{2-} , SO_3^{2-} and $S_2O_3^{2-}$ ions. Their content can be determined by redox titration under different conditions.

1. To a 20.00 mL sample containing S^{2-} , SO_3^{2-} and $S_2O_3^{2-}$ an excess of $ZnCO_3$ suspended in water was added. Upon completion of the reaction the solution was filtered into a 50.00 mL volumetric flask and diluted to the mark. To 20.00 mL of the filtrate an excess of aqueous formaldehyde was added. The mixture was acidified with acetic acid and titrated with 5.20 mL of 0.01000 M standard solution of iodine.

- Write down the net ionic equations of the reactions taking place during the analysis.
- Which ion, S^{2-} , SO_3^{2-} or $S_2O_3^{2-}$, can be determined by this method?
- Calculate the concentration of this ion in ppm in the initial solution.

2. A 20.00 mL sample of the 0.01000 M iodine solution was acidified with acetic acid and then combined with 15.00 mL of the filtrate above. The mixture was titrated with 6.43 mL of the 0.01000 M sodium thiosulfate standard solution.

- Write down the net ionic equations of the reactions taking place during the analysis.
- Which ion, S^{2-} , SO_3^{2-} or $S_2O_3^{2-}$, can be determined by this method taking into account the result of the previous experiment?
- Calculate the concentration of this ion in ppm in the initial solution.

3. A 10.00 mL sample of 0.05000 M iodine solution was acidified with acetic acid and then 10.00 mL of the original sample containing S^{2-} , SO_3^{2-} and $S_2O_3^{2-}$ were added. The mixture was titrated with 4.12 mL of 0.05000 M sodium thiosulfate standard solution.

- a) Write down the net ionic equations of the reactions taking place during the analysis.
- b) Which ion, S^{2-} , SO_3^{2-} or $S_2O_3^{2-}$, can be determined by this method taking into account the results of two previous determinations?
- c) Calculate the concentration of this ion in ppm in the initial solution.

Problem 14. MAGNESIUM DETERMINATION

To determine the amount of magnesium in a solution, a sample of the liquid was first acidified with HCl, then made slightly alkaline by addition of NH_3 and then combined with an excess $(NH_4)_2HPO_4$ in water. The precipitate of $MgNH_4PO_4$ formed was filtered off, washed with diluted aqueous NH_3 , annealed at $1000\text{ }^\circ\text{C}$ to constant mass and weighed.

Answer the following questions using numerical data given in the end of the text whenever necessary.

1. Write down the net ionic equation for the precipitation reaction taking place in course of the analysis.
2. Write down the equation for the reaction taking place in the course of annealing.
3. When determining the content of magnesium in a granulated medicine preparation calmagin 0.1532 g of the annealed precipitate were obtained from a 1.8005 g sample of calmagin. Calculate the mass percent of MgO in the preparation.
4. During the precipitation of $MgNH_4PO_4$ some impurities may coprecipitate such as $MgHPO_4$, $Mg(NH_4)_4(PO_4)_2$, $Mg_3(PO_4)_2$, $Mg(OH)_2$, $(NH_4)_2HPO_4$ and NH_4Cl . Some of these substances can undergo thermal decomposition at annealing. Write down the equations of the corresponding reactions.

5. Indicate if the presence of the impurities listed in Table below can lead to an error in the magnesium content as determined by the method described above. Put 0 in the Table if no error is expected, plus or minus sign if the error will be positive or negative respectively.

| Impurity | Error |
|---|-------|
| MgHPO ₄ | |
| Mg(NH ₄) ₄ (PO ₄) ₂ | |
| Mg ₃ (PO ₄) ₂ | |
| Mg(OH) ₂ | |
| (NH ₄) ₂ HPO ₄ | |
| NH ₄ Cl | |

6. At what maximum pH value the precipitation of MgNH₄PO₄ may be carried out to avoid simultaneous precipitation of Mg(OH)₂? Assume that the volume of the original sample was 200 mL and the content of magnesium in it was 0.10 g.

7. To determine the solubility product (K_{sp}) of MgNH₄PO₄ a NaOH solution was added dropwise until the beginning of precipitation to a 100 mL of solution containing 0.010 M MgCl₂, NH₄Cl and NaH₂PO₄ each. The precipitation started at pH 6.48. Calculate K_{sp} . Neglect the volume change during the experiment.

Reference data

| | | | |
|--------------------------------|--------------------|----------|----------------------|
| H ₃ PO ₄ | acidity constant | K_{a1} | $7.1 \cdot 10^{-3}$ |
| | | K_{a2} | $6.2 \cdot 10^{-8}$ |
| | | K_{a3} | $5.0 \cdot 10^{-13}$ |
| NH ₃ | basicity constant | K_b | $1.8 \cdot 10^{-5}$ |
| Mg(OH) ₂ | solubility product | K_{sp} | $6.0 \cdot 10^{-10}$ |
| H ₂ O | ionic product | K_w | $1.0 \cdot 10^{-14}$ |

Problem 15. INORGANIC PHOSPHATES: FROM SOLUTION TO CRYSTALS

Inorganic acids containing phosphorus and oxygen and most of the salts of these acids are composed of oxygen tetrahedra, each with the phosphorus atom in the center. The tetrahedra can either be isolated or share an oxygen atom so being linked by means of P–O–P bridges.

1. a) Draw the structure of the anions present in the neutral salts of the following acids: H_3PO_4 , H_3PO_3 , H_3PO_2 .

b) For the series of acids above, reveal the trends in:

- 1) acidity of the substances (compare the values of $\text{p}K_{a1}$),
- 2) O–P–O valence angle.

2. The formula of metaphosphoric acid can be written as $(\text{HPO}_3)_n$. This acid is composed of the phosphorus-oxygen tetrahedra either. Suggest the structure of this compound assuming the minimal number of phosphorus atoms in its molecule.

3. a) To estimate the relative charge of atoms in $\text{P}_n\text{O}_k^{(2k-5n)-}$ anion, let us define a special secondary parameter A_i of an atom i as the oxidation number of this atom, Z_i , divided by its coordination number, CN_i :

$$A_i = \frac{Z_i}{\text{CN}_i}.$$

The sum of the oxidation number (Z_N) of an atom N (for instance, phosphorus atom) and A_i values for the atoms forming the coordination environment (for instance, oxygen atoms) of the atom N gives the relative charge $Q(N)$ of the atom N :

$$Q(N) = Z_N + \sum_{i=1}^k \frac{Z_i}{\text{CN}_i}.$$

Calculate $Q_m(\text{P})$ for the PO_4 tetrahedron considering $m = 1, 2, 3$ and 4 of its oxygen atoms being shared with neighboring PO_4 -tetrahedra.

b) Perform similar calculations for TO_4 -tetrahedra linked through the common vertices, where

1) $T = \text{Si}$,

2) $T = \text{S}$.

4. Let us suppose that a tetrahedron with the minimal absolute value of $Q_m(\text{P})$ is the most stable towards hydrolysis.

a) Which value of m corresponds to the phosphorus-oxygen tetrahedron the most stable towards hydrolysis?

b) Which value of m corresponds to the TO_4 tetrahedron ($T = \text{Si}, \text{S}$) the most stable towards hydrolysis?

5. Isolated phosphorus-oxygen tetrahedra (without P-O-P bonding) can be found in crystalline substances. Mixed phosphates (V) M_aPO_b are known to be composed of PO_4 - and MO_4 -tetrahedra with each oxygen atom having the same number of M and P atoms coordinated to it.

a) Determine the $Q(\text{O})$ value for such compounds.

b) Suggest possible empirical formulas for such compounds.

6. Fluorapatite $\text{Ca}_5(\text{PO}_4)_3\text{F}$ is a constituent of human teeth. It can be synthesized using a double-diffusion method with a gelatin membrane separating solutions containing F^- , HPO_4^{2-} , and Ca^{2+} ions. The synthesis leads to a hybrid material – bioorganic polymer/inorganic phosphate, resembling tooth (or bone) tissue.

a) Give a reasonable composition of two solutions placed on different sides of the gelatin membrane, that allow preparation of fluorapatite as the target substance in this double-diffusion experiment.

| | 5 mM $\text{Ca}(\text{NO}_3)_2$ | 1 mM NaF | 3 mM Na_2HPO_4 |
|------------|---------------------------------|----------|--------------------------------|
| Solution 1 | | | |
| Solution 2 | | | |

b) Write down the balanced equation of the reaction described above leading to fluorapatite.

c) Calculate the osmotic pressure acting on the membrane at the beginning of this experiment (25 °C, activity of all ions is equal to 1).

Problem 16. FRUITS, VEGETABLES, ATOMS

When solving this problem none of the fruits or vegetables was destroyed!

In 1611 German mathematician and astronomer Johannes Kepler observed the stacking of cannonballs in a pyramid. He asserted there is the only way to fill the space the tightest possible with equal hard spheres, "...so that in no other arrangement could more pellets be stuffed into the same container". He was the first to formulate such a problem termed later as Kepler Conjecture. In 1998 Professor Thomas Hales¹ announced a solution to the Kepler Conjecture, which was published in a series of papers in "Discrete and Computational Geometry" starting from 1997. He considered 150 more variants of space filling besides that asserted by Kepler. Hales' solution required about 250 pages in a printed version and a size of 3 Gb in computer files. Thus, the term of close-packing of spheres (c.p.s.) widely accepted in the field of solid state chemistry passed through the rigorous mathematical proof and remained valid.

We do not request that you provide an alternative solution to this problem. However, you can check with our help how the basic law of space filling is applicable to our everyday life.

1. In order to avoid smashing tomatoes during their transportation, it is useful to arrange them on a shelf in a uniform single layer. Let us consider two types of packing (Fig. 2).

¹ Currently at the University of Pittsburgh, PA

a) Calculate the density of tomatoes packing (φ) for the case A and B as $\varphi = S_{\text{tomato}} / (S_{\text{void}} + S_{\text{tomato}})$.

b) Which type of the packing requires less shelf area?

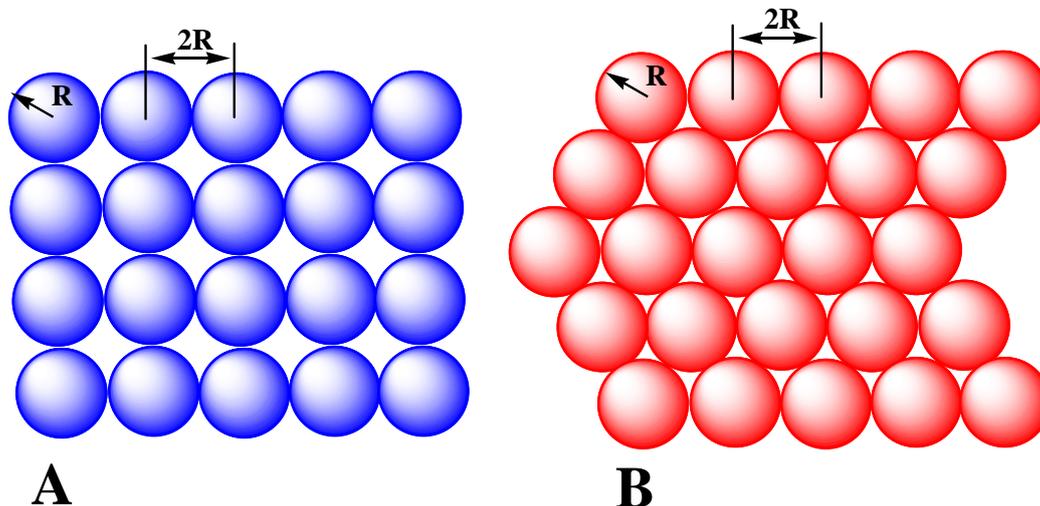


Fig. 2. Two possible types of packing tomatoes.

2. Hard vegetables such as potatoes or cabbage heads can be packed in containers.

Consider several types of packing:

(1) The first layer is of the type A (see Fig. 2). The second layer is an exact copy of the first, a vegetable in the second layer is above another one in the first layer (such a packing is termed usually as simple cubic packing, or s.c.).

(2) The first layer is of the type A. In the second layer each vegetable is above a void space in the first layer (body centered cubic packing, or b.c.c.).

(3) The first layer is of the type B. The second layer is an exact copy of the first, a vegetable in the second layer is above another one in the first layer (hexagonal packing, or h.p.).

(4) The first layer is of the type B. In the second layer each vegetable is above a void space in the first layer (hexagonal close packing, or h.c.p.).

a) Calculate the densities of packing for the cases (1) – (4).

b) Which type of packing is more efficient in the sense of van filling?

c) There are two alternatives to arrange the third layer in the case B: i) by placing vegetables right above the vegetables of the first layer (that is to place them into the voids of the second layer) or ii) by arranging vegetables right above the voids of the first layer (see the case B in Fig. 2). Calculate the density of packing φ for the second alternative which is called the face centered cubic packing – f.c.c.

d) A farmer filled the third layer in the way of f.c.c. and now can not figure out where the voids and vegetables of the first layer are. How does the value of φ vary due to the faults in regular sequence of closed packed layers?

3. Assume now that the enterprising farmer decided to place peaches into the van with watermelons. His bright idea was to place peaches into the voids of watermelon packing.

a) Estimate the maximal value of the $R_{\text{peach}} / R_{\text{watermelon}}$ peach/watermelon radii ratio that allows to avoid peach smashing in cases of:

- (1) cubic void within s.c.
- (2) octahedral void within b.c.c.
- (3) octahedral void within f.c.c.

b) How many peaches (maximum) per one watermelon can the farmer place using c.s., h.c.p., b.c.c. and f.c.c. types of packing?

c) What is the maximal φ value for c.s., b.c.c. and f.c.c. packings containing peaches in voids?

4. The fruits can go bad due to insufficient ventilation in the van.

a) In order to keep the voids in b.c.c. and f.c.c. packings the go-ahead farmer decided to put peaches only in the octahedral voids which are not linked by edges and faces. How many peaches per one watermelon can be packed in this case?

b) The enterprising farmer has got another idea: to fill all the octahedral voids in f.c.c. with peaches (you know about it), whereas (it's brilliant!) the tetrahedral voids with apples. How many apples per one watermelon can he arranged in this way?

Nature invents puzzles like the Kepler Conjecture. Opal is a natural stone composed of c.p.s.-packed SiO_2 microspheres. The main feature of opal is the distinguished shining (the so-called iridescence) when it is illuminated. This phenomenon is explained by the diffraction of visible light in accordance with Bragg's law:

$$\lambda = 2d \sin \theta$$

where λ is the wavelength of light, d is the distance between layers in c.p.s. of opal, 2θ is the angle between incident and diffracted beams (or, in other words, θ the inclination angle of the beam with respect to the surface of opal stone).

Opal is a prototype of photonic crystals, materials composed by closely packed microspheres with high refraction index. Optical spectra of photonic crystals demonstrate unusual features, for instance, photonic band gap (like electron band gap in semiconductors). Photonic crystals are considered to be the main active elements in photonics, the information technology of the future.

5. a) Find the minimal values of Miller indices – $(h k l)$ related to the first “permitted” reflection in f.c.c.
- b) Calculate the wavelength of light if the first reflection is observed for $2\theta = 60^\circ$. The radius of SiO_2 microspheres is equal to 450 nm. The dispersion of SiO_2 refraction index (that is, its dependence on wavelength) can be neglected.

Problem 17. CHAMELEONIC COBALT

Information was always regarded as the most valuable product resulting from mankind activity. It is not striking that recognition of this fact was followed by numerous efforts aimed at information safety. Cryptography seemed to be a convenient way to reach such safety from unrecorded time. Cryptography cannot be detached from sympathetic ink that becomes visible only after special treatment, for instance, heating. History knows a number of recipes of such ink, among them that based on salts of cobalt(II). Being pale-pink in color, cobalt ink is virtually invisible when dried on paper. However, once heated

with a candle flame, a letter written with such ink reveals hidden text colored in bright-blue.

We know other applications of cobalt(II) salts, less secret, but dependent on the color transition described above. Blue granules of silica-gel doped with Co(II) salt and placed into a desiccators to dry some product, become pink at last. This is the signal to regenerate silica-gel (just to dry, since it accumulates too much water). Similarly, a paper soaked with saturated solution of CoCl_2 turns blue in dry air due to formation of $\text{CoCl}_2 \cdot 4\text{H}_2\text{O}$, and changes its color back to pink $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$ in a humid environment. Apparently, the paper works as a humidity meter, hygrometer.

1. Using the thermodynamic data below, determine the threshold of air humidity (in %) specific to the response of such a hygrometer.

| Compound | $-\Delta_f H_{298}^\circ$, kJ mol ⁻¹ | S_{298}° , J mol ⁻¹ K ⁻¹ |
|---|--|---|
| $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}_{(\text{cr})}$ | 2113.0 | 346.0 |
| $\text{CoCl}_2 \cdot 4\text{H}_2\text{O}_{(\text{cr})}$ | 1538.6 | 211.4 |
| $\text{H}_2\text{O}_{(\text{liq})}$ | 285.8 | 70.1 |
| $\text{H}_2\text{O}_{(\text{g})}$ | 241.8 | 188.7 |

The “pink (sometimes, violet) ↔ blue” color transition described above is related to the reconstruction of the coordination sphere of Co^{2+} ion: octahedron ↔ tetrahedron. The examples discussed in a previous section deal with the transition $[\text{Co}(\text{H}_2\text{O})_6]_{\text{oct}}^{2+} \leftrightarrow [\text{Co}(\text{H}_2\text{O})_4]_{\text{tet}}^{2+}$. As a rule, coordination compounds with tetrahedral geometry are less abundant compared to octahedral ones. However, in particular case of Co^{2+} tetrahedral complexes competes with octahedral compounds.

2. To understand the reason behind such behavior, consider the following octahedral and tetrahedral complexes:

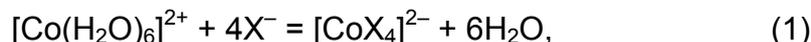
- a) $[\text{Cr}(\text{H}_2\text{O})_6]^{3+}$ and $[\text{Cr}(\text{H}_2\text{O})_4]^{3+}$,
- b) $[\text{Co}(\text{H}_2\text{O})_6]^{2+}$ and $[\text{Co}(\text{H}_2\text{O})_4]^{2+}$.

Draw diagrams for the case of an octahedral and a tetrahedral ligand field showing clearly the energy levels of all metal 3d-orbitals; indicate the d-orbital splitting parameter Δ . For each of the ions above use the appropriate diagram and fill it in with the electrons

available in the metal d -subshell. Calculate the Crystal Field Stabilization Energy (CFSE) for each of the ions.

Compare the results and draw a conclusion.

3. The following reaction



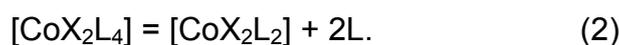
where $\text{X}^- = \text{Cl}^-$, Br^- , I^- , SCN^- , is used in some textbooks to illustrate Le Chatelier's principle related to equilibrium shifting. If one adds an excess of salt containing X^- , the solution becomes blue, and under dilution with water it turns back pale-pink.

a) Predict the signs of the enthalpy ($\Delta_r H_{298}^\circ$) and entropy ($\Delta_r S_{298}^\circ$) changes for the reaction (1).

b) What effect does temperature produce on the equilibrium (1)?

c) Consider reaction (1) and KCl and KSCN as a source of ions X^- for it. Which salt present in the same molar concentration shifts the equilibrium (1) to the right in a greater extent? Explain using the principle of Hard and Soft Acids and Bases (HSAB).

4. Consider a similar equilibrium (2):



a) If $\text{L} = \text{pyridine (py)}$, which ligand X (Cl^- or I^-) helps better shift the equilibrium (2) to the right? Explain using the principle of Hard and Soft Acids and Bases (HSAB).

b) If $\text{L} = \text{PH}_3$, which ligand X (Cl^- or I^-) helps better shift the equilibrium (2) to the right? Explain using the HSAB principle.

c) The coordination compound with the formula $[\text{CoX}_2\text{L}_2]$, where $\text{L} = \text{py}$, $\text{X} = \text{Cl}^-$ exists in two forms colored blue and violet. The structure of the former is quite apparent, whereas that of the latter is less obvious. For the violet form, draw a fragment

of its structure large enough to show clearly the coordination mode of the cobalt ion.

With some knowledge of coordination chemistry of Co(II) described above, you may be able to account for the transformations described below.

NaOH solution is added dropwise to a solution of Co(II) under cooling (0 °C), which results in a precipitate of blue color. If the precipitate is left at room temperature (25 °C) for a while, it becomes pink. If an excess of alkali is further added to the precipitate, it dissolves giving blue solution.

5. Write down equations corresponding to the transformations described above.

Problem 18. THE FORMOSE REACTION

Aldehydes have a high and versatile reactivity serving as indispensable reagents in the organic synthesis. Carbon atom of the carbonyl group is an electrophilic center. In the aldol condensation reactions a nucleophilic enol (or enolate) attacks the electrophilic carbonyl group of the other aldehyde (or ketone) molecule.

1. Fill in blank boxes in the representative aldol condensation reaction, and mark by letters **E** or **N** the respective nucleophilic and electrophilic reaction centers which take part in the process



The aldehydes lacking α -hydrogen atoms are commonly believed to be unable to take part in the aldol reactions as a nucleophilic component, thus such aldehydes are apparently unable to undergo self-condensation.

2. Such aldehydes are commonly referred to as *non-enolizable*. Why? Give any three examples of such aldehydes.

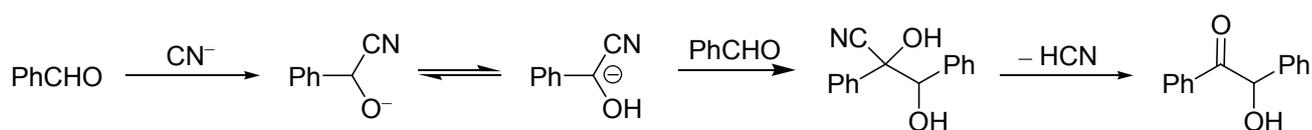
Formaldehyde is the most famous among such aldehydes. It was discovered by one of the founding fathers of organic chemistry, Alexander M. Butlerov as early as in 1859. Studying the compound Butlerov discovered a very interesting transformation of aqueous formaldehyde in the presence of lime into sugary syrup. The other great chemist Emil Fischer studied this transformation in more detail about half a century later, and discovered that a complex mixture of racemic carbohydrates is actually formed.



The mixture was given a name “*formose*”; the transformation since then is called *the formose reaction*. This reaction is very interesting due to its possible role in the generation of sugar molecules in a prebiotic Earth. Also it is quite promising from a practical viewpoint as a very inexpensive source of sugars for biotechnology given that formaldehyde is a readily accessible raw material which is produced in huge amounts from carbon and water.

3. Suggest a method for industrial preparation of formaldehyde from coal and water in no more than 3 stages.

The way formaldehyde enters the condensation remained an enigma for a long time since Fischer’s works. One of the possible keys to this problem is the so-called *Umpolung*². The essence of this important synthetic notion can be illustrated using the benzoin condensation as an example:

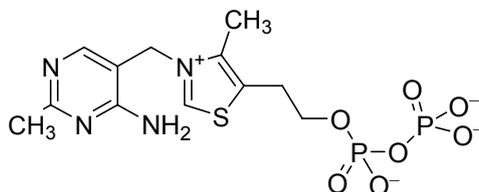


4. Mark in structure of the product (benzoin) the fragments coming from benzaldehyde and put the letters **E** and **N** over electrophilic and nucleophilic centers.

The intermediate generation of a nucleophilic reagent from a compound ordinarily behaving as an electrophile (or vice versa) is referred to as the *Umpolung* principle in modern organic chemistry.

² This German word is not usually translated in English due to lack of adequate and short translations.

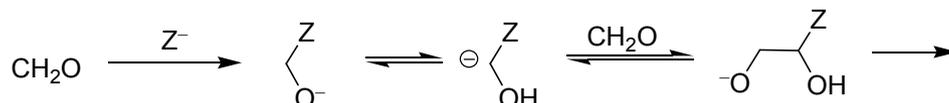
In order to avoid handling deadly cyanides, other compounds having similar CH-acidity, thiazolium salts, can be used. Such a non-trivial choice comes from a far-reaching analogy. One of such thiazolium salts, vitamin B₁ derivative, or thiamine pyrophosphate, is employed by Nature as a co-factor for trans-ketolases, that perform *in vivo* reactions closely resembling the benzoin condensation by transferring a carboxylic acid residue (acyl) as a nucleophilic rather than electrophilic reagent.



5. Mark in thiazolium the CH-acidic center equivalent to that in HCN. Draw the structure of the respective carbanion and show its resonance structures that account for the enhanced CH-acidity.

6. Alcohol addicts often suffer from an acute B₁ deficiency. Why?

A model of formose reaction has been studied. Formaldehyde in the presence of calcium hydroxide and vitamin B₁ (denoted as HZ in the Scheme below) gives the simplest ketotriose (dihydroxyacetone, DHA) in good yield.



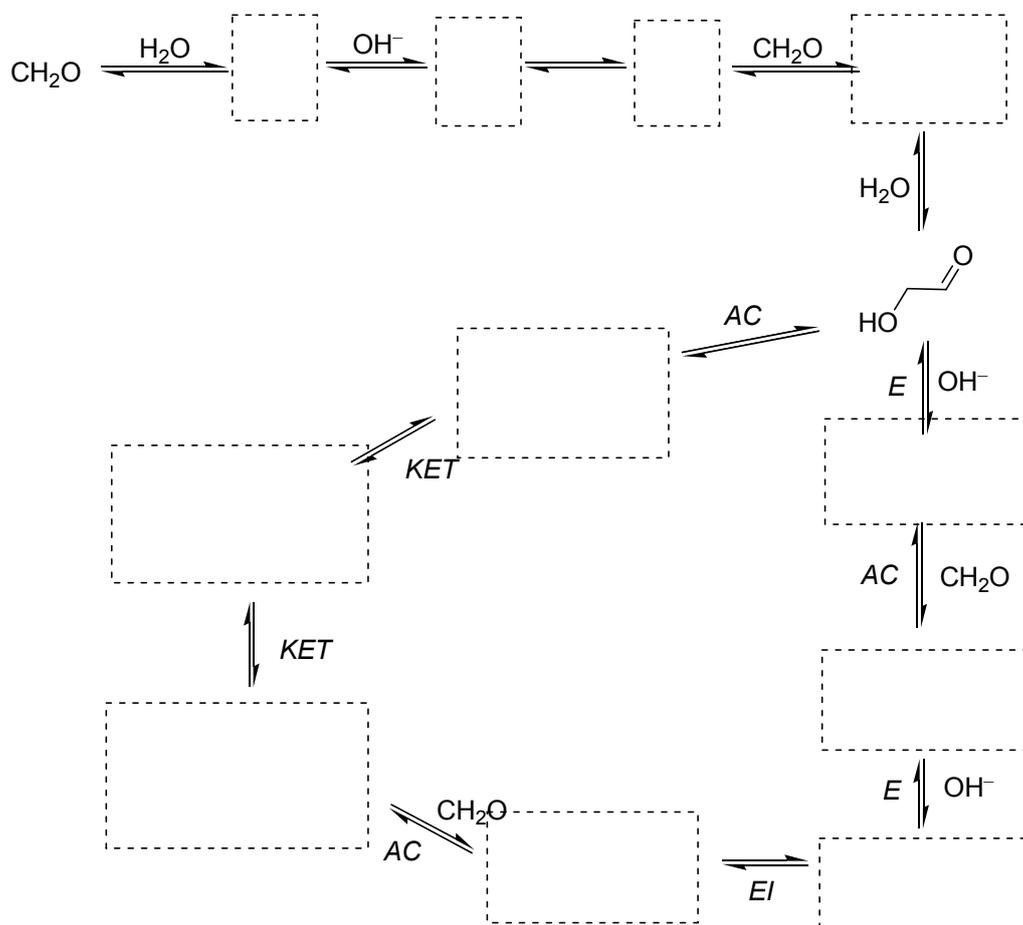
7. Complete this scheme to the final product.

With all these data at hand, we can try to crack the enigma of the real formose reaction. An essential clue is that the reaction of pure aqueous formaldehyde in the presence of lime is autocatalytic, which means that it is extremely slow at the beginning (there is an induction period), but once it starts it runs at an increasing rate until exhaustion of formaldehyde. Traces of *any* carbohydrate dramatically accelerate the reaction and immediately launch it if introduced within the induction period. The process involves a catalytic cycle consisting of aldol condensations (AC), keto-enol tautomerizations (KET), proton transfers leading to enolates (E), enolate or enol isomerizations (EI).

8. Fill in empty boxes on the simplified scheme of formose reaction below.

9. Show the step(s) involved in the induction period.

10. Show the catalytic cycle. What compound(s) serve(s) as catalyst(s)?



Problem 19. THE ANALOGY IN ORGANIC CHEMISTRY

Though not strict but rather an intuitive concept, the analogy (structural, electronic, stereochemical) is widely used by chemists for reasoning. For example, organic chemists often predict new reagents or even reactions by analogy with known ones.

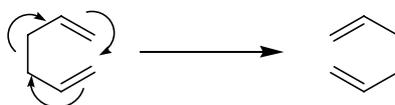
An important sort of analogy is heteroanalogy – the similarity of compounds or reactions differing by substitution of an atom or group by another atom or group having the same type of bonding.

Thus, heteroanalogues of aldehydes are iminium salts, e.g. a well-known Eschenmoser's salt $\text{CH}_2=\text{NMe}_2^+\text{I}^-$.

1. Which type of reagent is the cation of Eschenmoser's salt? Electrophile , nucleophile , free radical , Lewis acid , oxidant , protecting group

2. Write by analogy the reaction of Eschenmoser's salt with acetone. Why does this reaction not require a catalyst?

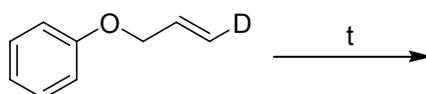
Further we may consider a heteroanalogy concept with respect to reactions. E.g. there is the Cope rearrangement, which takes place if 1,5-dienes are being heated. The reaction is a concerted movement of 6 electrons to involve two π -bonds and a σ -bond, a sigmatropic shift.



3. What products are formed on prolonged heating of 1,5-hexadiene substituted at C1 with one deuterium atom in inert atmosphere (possible isotope effects are to be neglected)?

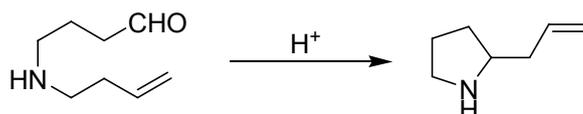
If we take vinyl allyl ether $\text{CH}_2=\text{CH}-\text{O}-\text{CH}_2\text{CH}=\text{CH}_2$ in place of diene, the same sort of rearrangement takes place, but with a more interesting result leading to a compound of the other class, unsaturated ketone. Such hetero- (oxa-)analogue is usually called the oxo-Cope rearrangement, or Claisen rearrangement. This reaction was discovered by a happy chance by great German chemist Ludwig Claisen.

4. Complete the reaction



The rearrangements of this sort are interesting because new reactive groups can form in a very simple process, and these newly-born groups can enter further reactions in the same reaction mixture without the isolation of intermediate compounds. Such chains of transformations are often called the domino-reactions, by analogy with a well-known trick when a long chain of standing dominoes is made to fall by a single click.

5. Your task would be to imagine how the following domino-process, which is initiated by a drop of strong acid and a dehydrating agent, such as $\text{HC}(\text{OEt})_3$, takes place



Write the steps involved in this process.

Problem 20. KETO-ENOL TAUTOMERISM

Aqueous or alcoholic solutions of ketones or aldehydes can be titrated by solutions of halogens or interhalides. In order to obtain reproducible results, the titration should be performed fast in the presence of buffer salts, such as NaHCO_3 .

Thus, to 10 g of cyclohexanone in aqueous methanol were added 2.00 mmol NaHCO_3 , and 1.00 ml 2.00 N methanolic solution of ICl . After thorough mixing an excess of aqueous NaI solution was added, followed by titration by 1.594 ml of 1.00 N $\text{Na}_2\text{S}_2\text{O}_3$ using starch as indicator.

1. Write the reactions involved in the analysis.
2. What compound reacts with ICl ? Estimate the content of this compound in cyclohexanone.
3. What is the role of buffer salt? What can happen if Na_2CO_3 is taken in place of NaHCO_3 ?

A colorless substance **A** with the empirical formula C_2H_2O shows in ^{13}C NMR only two signals at 94 and 159 ppm. The reactions of **A** with halogens or interhalides are instantaneous, but titration, as described above, is not useful as more than one mole halogen per mole **A** is consumed to give off heavy precipitates.

A readily reacts with aldehydes in the presence of either acidic or basic catalysts, to form products of 1:1, 1:2 or 1:3 stoichiometry (depending on reagent ratio). Such products are often colored, which is used in many well-known qualitative reactions for aldehyde-containing materials. For example, carbohydrates give red coloration when treated by **A** and a drop of HCl.

Under alkaline conditions **A** reacts with methyl iodide to give a mixture of products. With a large excess of MeI a single compound **B** is produced. **B** turned out to be identical to a known trimer of dimethylketene formed under the conditions of basic catalysis. On the other hand, if the reaction of **A** with excess MeI is performed in the presence of $NaHCO_3$ a different compound **C** is formed. This compound possesses a fine odor and has been identified as one of important constituents of rose flavor. In 1H NMR compound **B** shows a single resonance, while **C** shows two sharp singlets with integral intensities ratio of 1:3.

The reaction of **A** with $NaHSO_3$ on heating gives colorless water-soluble material (brutto-formula $C_6H_5NaO_5S$) showing a purple coloration with $FeCl_3$ solution. The ^{13}C NMR spectrum in D_2O shows 4 signals at 157, 144, 106, 105 ppm.

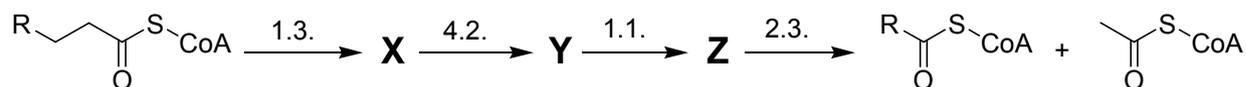
The reaction of **A** with hydroxylamine gives a compound **D** (brutto-formula C_2H_3NO), which is cleanly reduced by H_2 over Raney-Ni catalyst to give a compound **E** (brutto-formula C_2H_3N) rapidly darkening in the air. The compound is poorly soluble in water, but readily dissolves in dilute HCl. Boiling of this solution gives back **A**.

4. Determine the structures of **A**, **B**, **C**, **D**, **E**.

5. Write the reactions mentioned in the text

Problem 21. UNUSUAL PATHWAYS OF FATTY ACID OXIDATION: ALPHA-OXIDATION

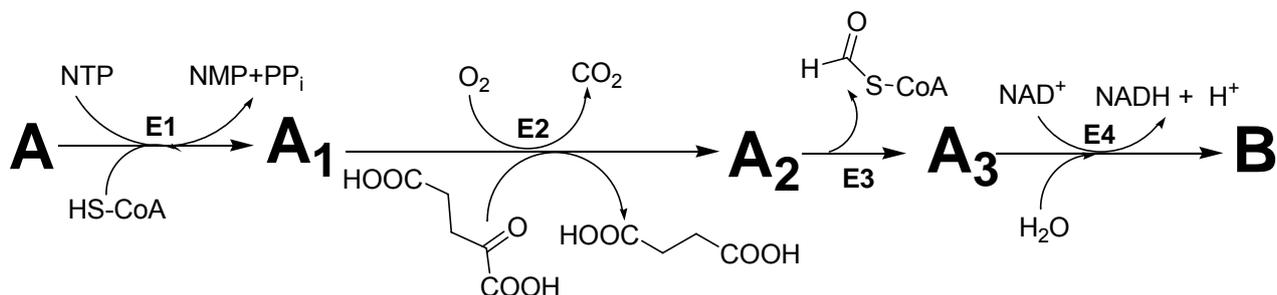
Oxidative destruction of fatty acids is a universal biochemical process inherent in all living systems. The so-called β -oxidation is the dominating pathway of fatty acid degradation in mitochondria. It can be described by the following scheme:



At all stages of β -oxidation, acyl residues are linked with coenzyme A by thioester bond. On the above scheme, classes and subclasses (numbers beyond the arrows) of enzymes catalyzing corresponding reactions are given in accordance with IUB classification. Note that substituent R remains unchanged within one cycle turnover.

1. Draw structures (without stereochemical details) of metabolites **X**, **Y** and **Z** using symbol "R" for the unchanged part of acyl residue.

Phytanic acid **A** is a saturated fatty acid which is found in nature as a mixture of two diastereomers. It is not involved in β -oxidation due to peculiar features of its structure. Nevertheless, mammals metabolize it into pristanic acid **B** with retention of configuration of chiral atoms. The latter process (usually referred to as α -oxidation) occurs in special cellular organelles, peroxisomes. Reaction equations on the scheme below illustrate metabolism of **A**:



NMP and NTP are mono- and triphosphates of ribonucleoside **N** (A, C, G or U), respectively, PP_i – pyrophosphate, CoA-SH – coenzyme A, NAD⁺ and NADH – oxidized and reduced forms of nicotine amide adenine dinucleotide, respectively, **E1-E4** – enzymes catalyzing corresponding reactions.

Biosynthesis of **A**₁ catalyzed by **E1** is a two-stage process. The intermediate formed contains phosphorus and oxygen in a molar ratio of 1:8.

2. From the list of reaction types given below, choose those which correspond to the stages catalyzed by **E1** and **E3**.

- Formation of an ester of ribonucleoside phosphate and carbonic acid,
- transfer of a phosphoric acid residue on a substrate due to cleavage of high energy bond of another substrate (kinase reaction),
- hydrolysis of an ester bond,
- formation of a thioester of carbonic acid,
- oxidative decarboxylation,
- cleavage of a carbon-carbon bond.

3. Draw the intermediate of the **E1** catalyzed reaction considering the formula of phytanic acid as R–COOH, where R is a hydrocarbon residue.

B is further metabolized in a number of consecutive cycles of β -oxidation. Data on oxidative destruction of pristanic acid are given in the table below.

| Stage | Cleavage Product(s) |
|---|--|
| Formation of pristanoyl CoA | No |
| The 1 st cycle of β -oxidation | Propionyl CoA |
| The 2 nd cycle of β -oxidation | Acetyl CoA |
| The 3 rd cycle of β -oxidation | Propionyl CoA |
| The 4 th cycle of β -oxidation | Acetyl CoA |
| The 5 th cycle of β -oxidation | Propionyl CoA |
| The 6 th cycle of β -oxidation | Acetyl CoA |
| The 7 th cycle of β -oxidation | Propionyl CoA + Formyl CoA (final products of degradation) |

4. Determine the empirical and molecular formulae of phytanic acid **A** without deciphering α -cycle and establishing structural formula of pristanic acid.

5. Draw structural formulae of **A** and **B** with stereochemical details. Take into account that all chiral centers in these fatty acids but that nearest to the carboxylic group exist in R-configuration only.

6. Explain why phytanic acid cannot be involved in β -oxidation.

The enzyme catalyzing the first reaction of β -oxidation cycle is stereospecific. Acyl CoA is transformed by this enzyme only in case the chiral center most distant from ω -carbon atom is in S-configuration. There exists a special enzyme, racemase AMCAR (marker of some oncologic pathologies), which transforms pristanic acid and some of its β -oxidation metabolites by catalyzing R \rightarrow S transition in the chiral center most distant from ω -carbon atoms.

7. Suggest the mechanism of pristanoyl CoA racemization.

8. Draw (with stereochemical details) those metabolites of pristanic acid oxidation which are AMCAR substrates.

During α -oxidation of **A** in mammals, only one pair of diastereomers is formed in **E2** catalyzed reaction.

9. Based on sterical considerations, suggest configuration (R or S) of chiral centers in diastereomers **A2**.

Problem 22. UNUSUAL PATHWAYS OF FATTY ACID OXIDATION: OMEGA- AND (OMEGA-1)-OXIDATION.

To be solved after problem 21

ω -Oxidation is one of metabolic pathways of fatty acids, though less common than β -oxidation. This unusual route starts with oxidation of the methyl group of a fatty acid to give new carboxyl group. The resulting dicarboxylic acid is further involved into several β -oxidation cycles developing in the direction towards the carboxyl group initially present in the acid. All reactions of ω -oxidation are non-stereospecific.

Due to peculiar features of its structure, synthetic saturated fatty acid **D** can be involved in mammals into ω -oxidation only (neither in α - nor in β -oxidation). The resulting dicar-

bonic acid **E** is metabolized into corresponding acyl CoA, which is further subjected to seven consecutive cycles of β -oxidation to give seven acetyl CoA molecules. The formula of the remaining metabolite **F1** of the pathway is $C_{27}H_{39}N_7P_3SO_{19}^{5-}$. **F1** exists as anion at physiological pH values. Its hydrolysis leads to two products, one of which, substance **F2**, does not contain chiral carbon atoms.



1. Draw the structures of compounds **D**, **E**, **F2** and anion **F1** at pH 7. Show evidence to prove that the answer is unambiguous.
2. Explain why fatty acid **D** cannot be involved in both α - and β -oxidation.
3. Propose the structure (without stereochemical details) of synthetic fatty acid **G**, an isomer of compound **D**, which contains the same number of carbon atoms in the main chain and cannot be involved in both α - and β -oxidation for structural reasons.

(ω -1)-oxidation is another pathway of fatty acid degradation in mammals. It plays an important role in metabolism of prostaglandins and development of several genetic diseases. One (ω -1)-oxidation cycle includes five two-electron oxidation reactions of a fatty acid.

Fatty monocarboxylic acid **H** that contains 75.97% C, 12.78% H, and 11.25% O by mass is widespread in nature. It gives compound **J** as the final product of (ω -1)-oxidation cycle. Compound **I** (72.42% C, 11.50% H, 16.08% O by mass) is one of intermediates of the pathway from **H** to **J**. 1H NMR spectrum of **I** contains two singlets with different integral intensities and a number of multiplets. Integral intensity of any multiplet differs from those of singlets. One of the singlets is characterized by the maximal integral intensity among all the signals in the spectrum.

4. Draw the structures of **H** and **I**. Show evidence to prove that the answer is unambiguous.
5. Determine how many steps of two-electron oxidation of **H** are required to produce **I**, if it is known that the entire ω -pathway is a part of (ω -1)-pathway.

6. Draw the structure of **J**.

α -Oxidation is impossible for patients with hereditary pathology Adult Refsum Disease (ARD) due to genetically determined absence of an enzyme of this oxidation pathway. Metabolism of phytanic acid **A** (a mixture of two diastereomers enriched with R-epimer, i.e. R>S, see problem 21) in organisms of such patients leads to dicarboxylic acid **C** (non-equivalent mixture of two enantiomers, R>S).

7. Determine how many steps of oxidation pathways given below are needed to obtain **C** from **A** in organisms of patients with ARD, if it is known that malonyl CoA is not released at the first β -oxidation cycle.

β -oxidation _____
 ω -oxidation _____
 $(\omega-1)$ -oxidation _____

AMCAR is the only epimerase involved in the process of oxidation of **A** to **C** (see problem 21 for detailed information on AMCAR).

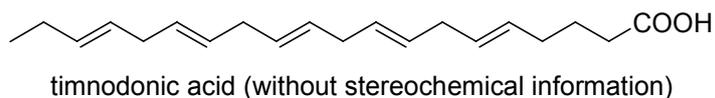
8. Draw formula(e) (with stereochemical details) of intermediate(s) of **A** oxidation in organisms of patients with ARD, that can be AMCAR substrates.

Problem 23. UNUSUAL PATHWAYS OF FATTY ACID OXIDATION: PEROXIDATION

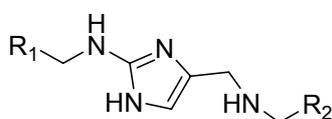
Peroxidation of lipids, in particular of those found in biomembranes and lipoproteins, is considered as an important stage in the development of numerous diseases including atherosclerosis. Lipids containing residues of polyunsaturated fatty acids (PUFA) are most liable to oxidation of this type.

X is one of the final products of peroxidation of any polyunsaturated acids in mammals. **X** can be also obtained by reductive ozonolysis of PUFA.

1. Write down the overall reaction of exhaustive ozonolysis of timnodonic acid with subsequent treatment of the reaction mixture with dimethyl sulfide.



X reveals high reaction ability towards various biomolecules including proteins. In particular, it interacts non-enzymatically with amino acid residues of albumin, an important transport protein of serum. As a result, side groups of two canonical amino acids are cross-linked. The linker formed in this reaction is depicted below (R_1 and R_2 are fragments of polypeptide chain of the protein):

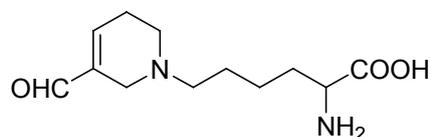


2. Draw (with stereochemical details) the structures of **X** and canonical amino acids, side groups of which are involved in the cross-linking.

3. Suggest mechanism of the linker formation, if it is known that only water molecules are released during the cross-linking.

Y is another product of peroxidation of lipids. It contains the same number of carbon atoms as **X** and interacts with both proteins and nucleic acids.

Interaction of **Y** with lysine residues present in a protein results in formation of residues of non-canonical amino acid N^ϵ -(3-formyl-3,4-dehydropiperidino) lysine (FDP-lysine):



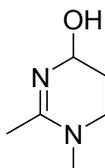
4. Draw the structure of **Y**, taking into account that equimolar amount of water is released upon FDP-lysine formation.

5. Suggest mechanism of formation of FDP-lysine residue if the starting lysine residue is a part of a protein. Note that Michael reaction is one of the steps of the pathway.

Interaction of **Y** with nucleoside **Z** found in nucleic acids results in an adduct, nucleoside **Z1**. Mass spectrum of **Z1** obtained by using fast atom bombardment mass spectrometry (FAB-MS) contains two major peaks corresponding to monoprotonated fragments ($M+H^+$), m/z values being equal to 191 and 307.

6. Draw the structure of **Z**, if its reaction with **Y** gives solely product **Z1**.

Z1 contains a base, a fragment of which is given below:



7. Draw the structure of **Z1**.

Problem 24. BIOLOGICALLY ACTIVE PEPTIDES AND THEIR METABOLIC PATHWAYS

(*Hint*: for calculations round all values of atomic masses of elements to integers)

Angiotensins (Ang) form a class of biologically active oligopeptides with numerous significant effects on human organism. They play an important role in regulating blood pressure, maintaining water-saline balance and performing intellectual and mnestic functions.

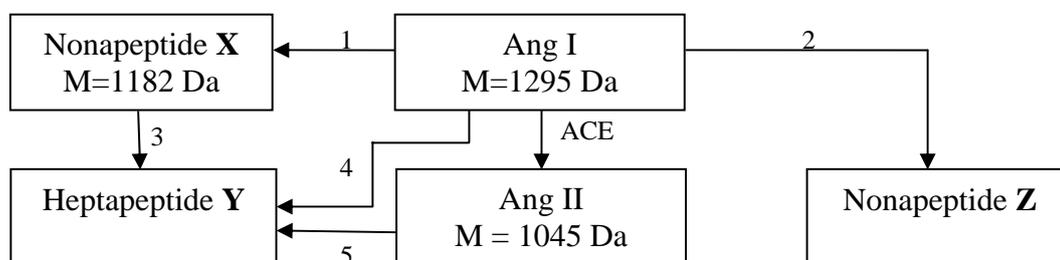
Decapeptide angiotensin I (Ang I) is the initial oligopeptide, a precursor of all members of the class. Complete acidic hydrolysis of Ang I leads to the mixture of nine amino acids: aspartic acid, arginine, valine, histidine, isoleucine, leucine, proline, tyrosine and phenylalanine.

Asparagine is hydrolyzed to form aspartic acid under the conditions required for complete hydrolysis of peptides.

1. Write down the equation of the acidic hydrolysis of asparagine.

Enzymes of several groups are involved in the metabolism of angiotensins. The first group includes amino peptidases (AMA and AMN), which cut off amino acids or peptide fragments from N-terminus of oligopeptides. The second group is represented by carboxypeptidases (Angiotensin-converting enzyme, ACE and its homolog ACE2), which cut off amino acids or peptide fragments from C-terminus of oligopeptides. The third group includes peptidases (neutral endopeptidase (NEP) and prolyl endopeptidase (PEP)), which split peptide bonds formed by specific amino acids residues.

Ang I is metabolized in man according to the scheme below:



1-5 are peptidases catalyzing corresponding reactions. Each of these peptidases catalyzes hydrolysis of only one peptide bond. One and the same peptidase may be encoded by different numbers.

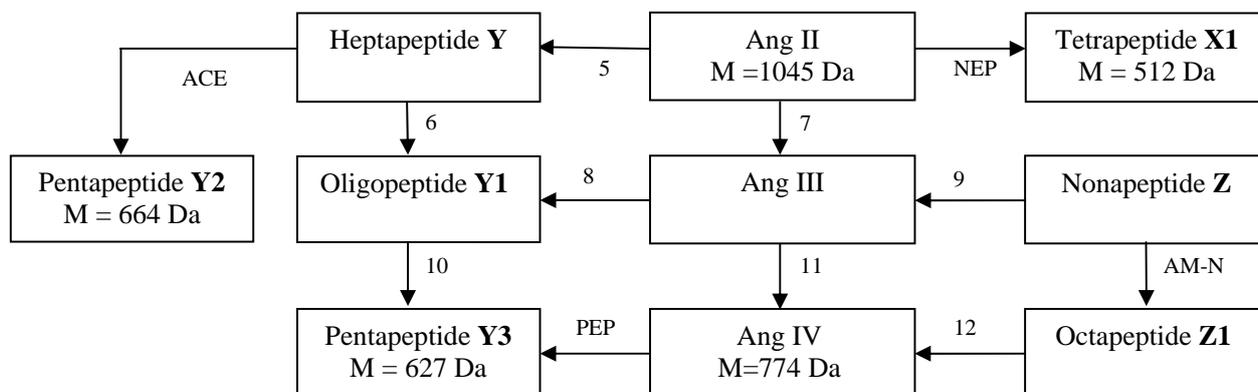
To name angiotensins, a special nomenclature has been developed. Amino acid residues of Ang I are enumerated from N- to C-termini. Since all angiotensins contain fragments of Ang I, the word «angiotensin» in their names is followed by Arabic numerals in parenthesis, indicating the positions of N- and C-terminal residues they occupied in Ang I. For instance, Ang I should be named according to the nomenclature as «angiotensin (1-10)».

2. Write down all possible variants of amino acids and/or oligopeptides, which can be cut off as a result of Ang II formation from Ang I.

3. Name oligopeptides X, Y and Z according to the Angiotensin nomenclature. Determine whether enzymes 1-3 are amino or carboxypeptidases.

4. Determine the gross amino acid content of Ang I. Show evidence to prove that the answer is unambiguous.

Metabolic pathways of Ang I derivatives are summarized in the following scheme:



6-12 are peptidases catalyzing corresponding reactions. One and the same peptidase may be encoded by different numbers.

Pancreatic proteinase trypsin catalyzes hydrolysis of peptide bonds formed by carboxyl groups of arginine or lysine. **Z1** has the highest molecular mass among all peptides formed as a result of trypsin catalyzed proteolysis of Ang I.

5. Determine which fragments are cut off as a result of the transformation from Ang II to Ang IV.

PEP selectively cleaves peptide bonds formed by carboxyl group of proline.

6. Determine the C-terminal amino acid in Ang II and structure of the dipeptide released when heptapeptide Y is treated with ACE.

Pancreatic proteinase chymotrypsin catalyzes hydrolysis of peptide bonds formed by carboxyl groups of aromatic amino acids phenylalanine, tyrosine or tryptophane. Quite often chymotrypsin also reveals specificity towards leucine, which is close to the mentioned above amino acids in hydrophobicity. Only two tetrapeptides are formed when Ang II is treated with chymotrypsin.

7. Write down the finally established exact amino acid sequence of Ang I.
8. Name oligopeptides **X1**, **Y1** and **Z1** according to the Angiotensin nomenclature.

Problem 25. RADICAL POLYMERIZATION

Radical polymerization is one of the most common methods of polymer synthesis. It involves the following stages:

Initiation – the stage at which active particles usually referred to as radicals appear as a result of particular chemical reaction and/or changes of physical properties of the system (heating, irradiation).

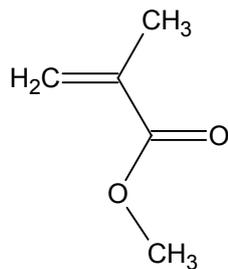
Chain propagation – consecutive addition of monomer molecules to a radical resulting in formation of new radicals of bigger size. Usually the rate constant of propagation is considered to be independent of polymerization degree of a growing radical (assumption of equal reactivity).

Chain termination – the stage at which chain growth is stopped due to bimolecular interaction of radicals. Recombination and disproportionation are possible ways of chain termination.

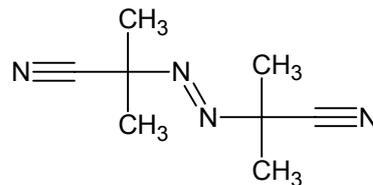
Chain transfer – the stage at which an inactive polymer molecule is formed due to interaction of a propagating radical with a chain transfer agent. This process is accompanied by transformation of the transfer agent into new radical. The latter can either initiate growth of a new polymer chain or terminate the chain. Molecules of the monomer, solvent or special additives can act as chain transfer agents.

To obtain poly-(methyl methacrylate) (poly-MMA), its monomer (9.4 g) was heated to 60 °C in the presence of 0.1 g of α,α' -azodiisobutyronitrile (AIBN) and 0.5 g of α -chlorotoluene. The density of the reaction mixture is 0.91 g/cm³. The rate constants of elementary stages are: $k_{in} = 7.2 \cdot 10^{-4} \text{ s}^{-1}$ (initiation), $k_p = 7.1 \cdot 10^2 \text{ l} \cdot \text{mol}^{-1} \cdot \text{s}^{-1}$ (propagation), $k_t = 2.6 \cdot 10^7 \text{ l} \cdot \text{mol}^{-1} \cdot \text{s}^{-1}$ (termination). Initiation efficiency is $f_{in} = 0.8$. Constants of chain transfer are: $C_A = 4.2 \cdot 10^{-4}$ (to α -chlorotoluene) and $C_M = 1.0 \cdot 10^{-5}$ (to the monomer).

Hint: chain transfer constant is defined as the ratio of the rate constants of chain transfer to a given species and chain propagation ($C = k_{tr} / k_p$).



MMA



AIBN

1. Write down reaction equations for initiation, chain propagation, chain termination, and chain transfer in the above given system.
2. Write down reaction equation(s) which decrease(s) initiation efficiency f_{in} .
3. Write rate equations for:
 - a) generation of active radicals
 - b) monomer consumption
 - c) changes of the concentration of radicals
4. Express equilibrium concentration of radicals under steady-state conditions as a function of kinetic parameters of elementary stages.
5. Express the rate of monomer consumption (rate of polymerization) as a function of immediate concentrations of the monomer and initiator and kinetic parameters of elementary stages. Find the order of polymerization reaction on the monomer and initiator.

Polymer obtained in the described above system at low conversion (less than 10% of the monomer consumed) possesses a number-average degree of polymerization P_n of 125.

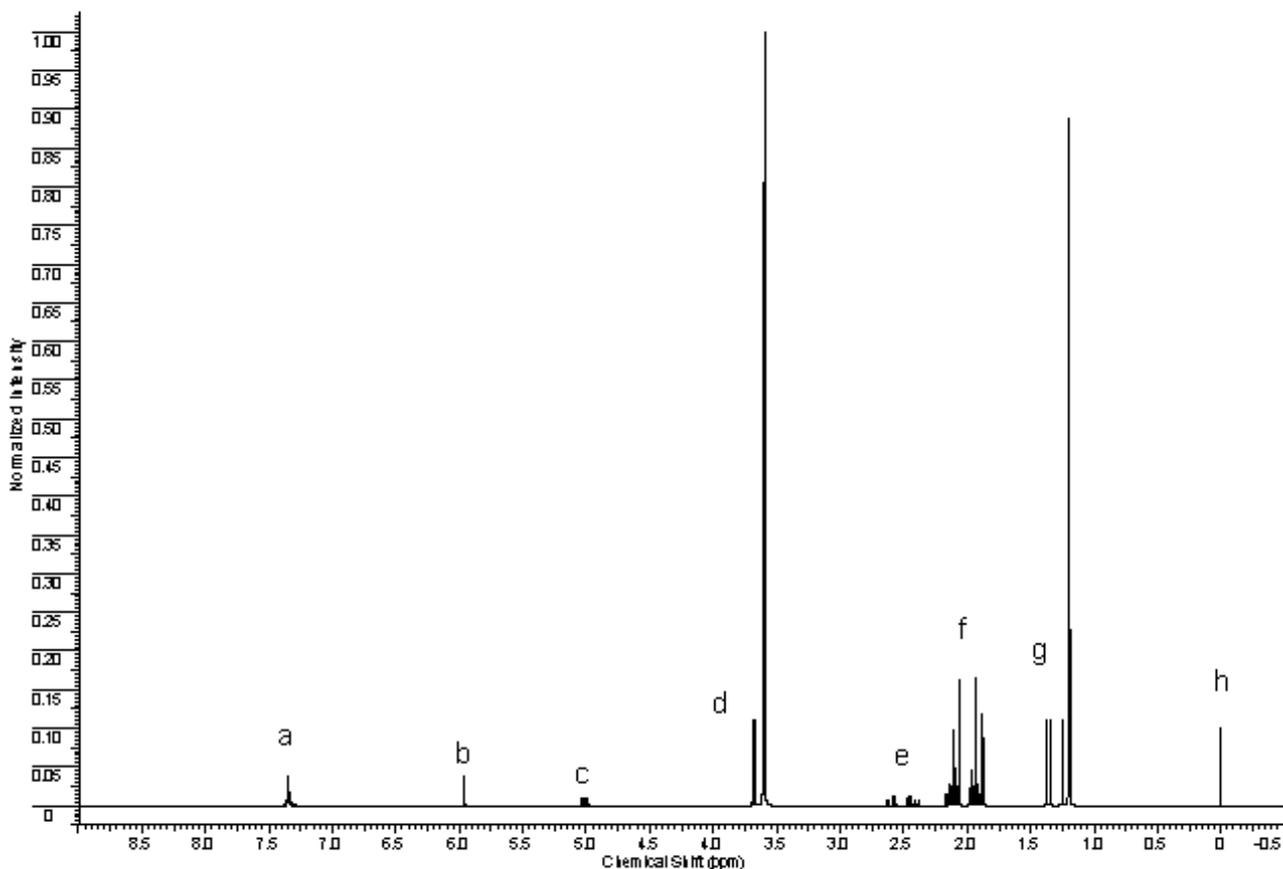
6. Determine the value of the rate constant of termination via disproportionation. Arrange the following processes in the decreasing order of their influence on P_n value.
 - a) chain termination
 - b) chain transfer to monomer

c) chain transfer to α -chlorotoluene

^1H NMR spectrum of a polymer obtained according to the above procedure is given hereunder.

7. Deduce the structure of the polymer using integral intensities of characteristic peaks given in the table.

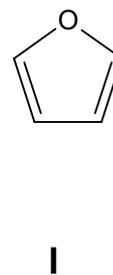
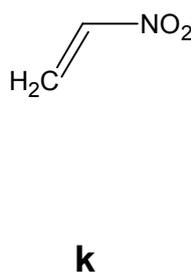
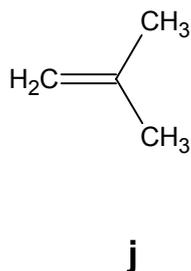
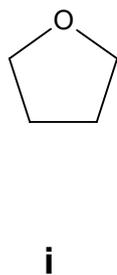
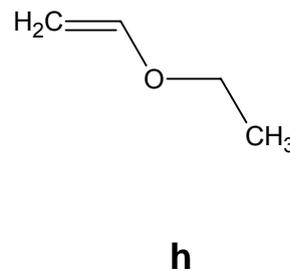
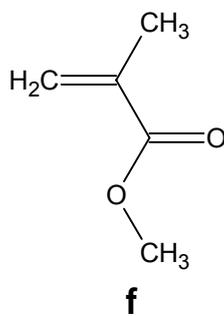
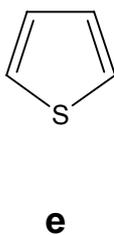
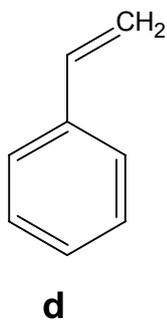
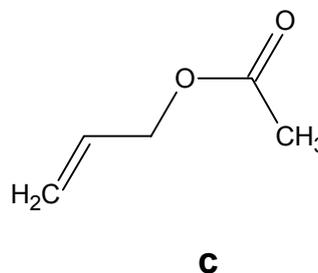
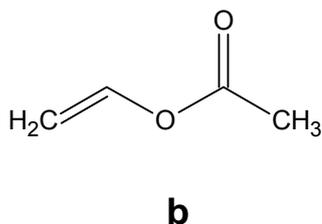
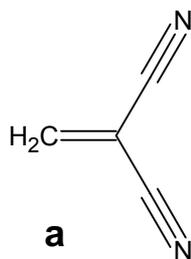
| Signal | Integral intensity |
|--------|--------------------|
| a | 5.0 |
| b | 1.0 |
| c | 1.0 |
| d | 42 |
| e | 2.0 |
| f | 27 |
| g | 39 |
| h | 4.5 |



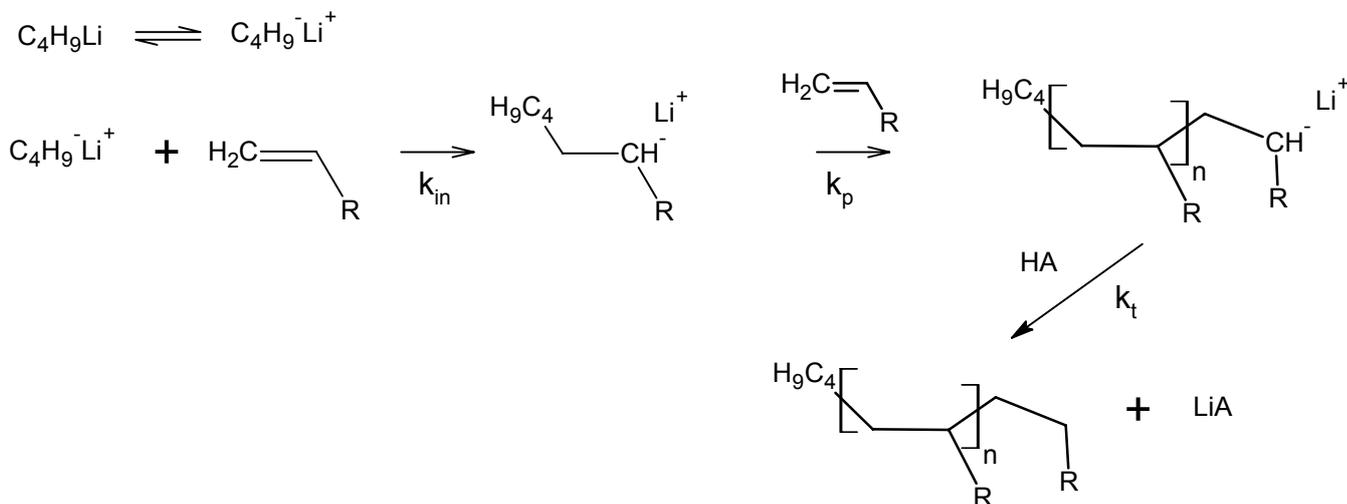
Problem 26. IONIC POLYMERIZATION

Polymerization may be initiated by ionic species. Depending on the charge on the end group of a propagating chain, cationic and anionic polymerization types are distinguished. Ionic as well as radical polymerization involves the stages of initiation, propagation, termination and chain transfer. Cationic polymerization is initiated by strong acids and other electrophilic compounds, whereas anionic by strong bases and electron donors.

1. For each monomer given below, choose polymerization type(s) (radical, anionic, cationic) which it can be involved in.



Anionic polymerization initiated by metal alkyls can be described by the following kinetic scheme, which includes stages of initiation, chain propagation and chain termination. The latter occurs as a result of carbanion reaction with a terminating agent, acid HA.



2. a) Write down the rate equation for monomer consumption, expressing concentrations of monomer and active chains (macroanions) as $[M]$ and $[M^-]$, respectively.
- b) Anionic polymerization allows synthesis of nearly monodisperse polymer. Based on this fact, compare qualitatively rate constants of initiation and chain propagation.
- c) Calculate molecular mass of the polymer obtained as a result of polymerization of 100 g of styrene in 600 ml of 1,4-dioxane in the presence of 0.234 g of naphthalene and 0.042 g of metallic sodium, if 58.9% of the monomer was consumed during polymerization.

Polymerization is a perspective approach towards design of chain molecules of various shape and size. Still chain termination can be regarded as a drawback of the method, since it leads to species not capable of attaching new monomer units.

3. a) What chain termination processes are probable for radical and anionic polymerization? Fill in the table.

| Type of chain termination | Radical polymerization | Anionic polymerization |
|---------------------------|------------------------|------------------------|
| Disproportionation | | |
| Recombination | | |
| Chain transfer to solvent | | |
| Chain transfer to monomer | | |

b) Explain why a polymer obtained by anionic polymerization has narrower molecular mass distribution than that obtained by radical polymerization.

c) The following solvents are used as a medium for anionic polymerization: (a) benzene; (b) 1,4-dioxane; (c) tetrahydrofuran; (d) 1,2-dimethoxyethane. Arrange the solvents in the order of increasing polymerization rate.

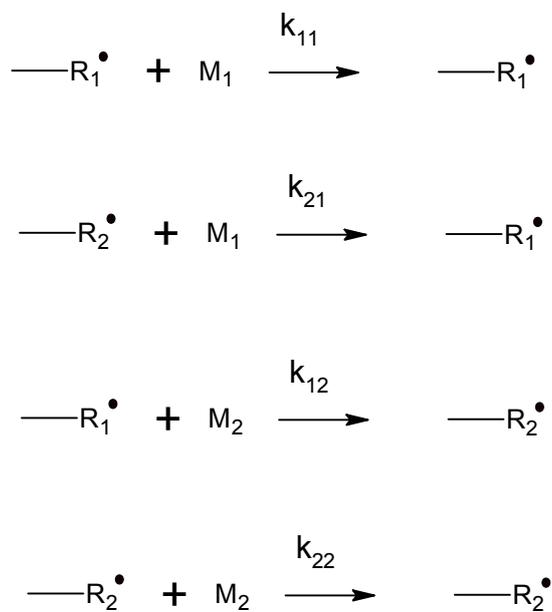
d) Compare the rates of anionic polymerization with sodium, potassium and cesium naphthalenides used as initiators.

Problem 27. CO-POLYMERIZATION

To synthesize macromolecules with complex architecture one can use various approaches: apply different types of polymerization, vary initiators, solvents and reaction conditions, copolymerize different monomers, as well as modify the obtained polymers. Some examples of copolymers are given in the table hereunder.

| Type of a copolymer | Schematic structure | Abbreviation |
|---------------------|---|----------------------------|
| Block | AAAAAAAAAAABBBBBBBBBBBBBBB | poly(A)-block-poly(B) |
| Alternating | ABABABABABABABAB | poly(A-alt-B), poly(AB) |
| Statistical | AABABAABBBAAABBBABAABABAAB | poly(A-stat-B) |
| Graft | <pre> AAAAAAAAAAAAAAAAAAAAAAAAAAAA B B B B B B B B B B B B B B B B B B </pre> | poly(A)-graft-poly(B) |
| Gradient | AAAAABAABAAABBABABBBBABB | poly(A-grad-B) |

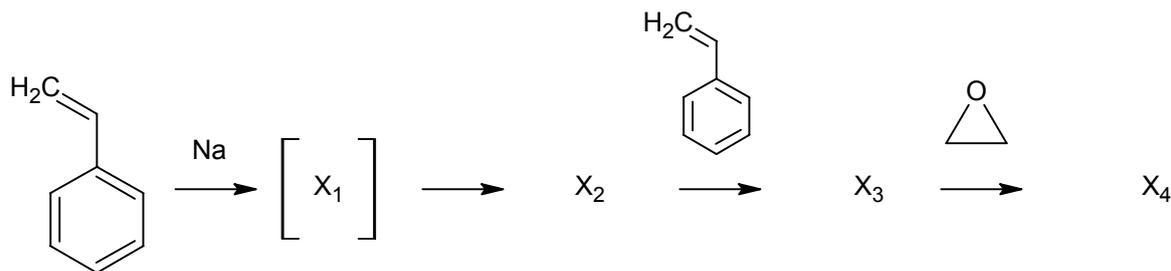
While developing copolymerization technique it is important to take into account relative reactivity of monomers. Kinetics of copolymerization can be described by a set of elementary reactions with corresponding rate constants. In the case of binary radical copolymerization four elementary reactions of chain propagation should be considered (end-unit model):



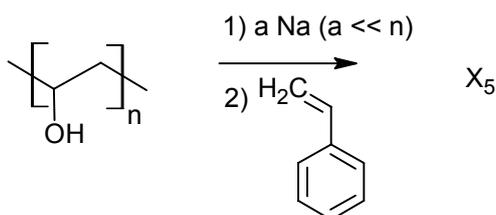
Relative reactivity of monomers in copolymerization is characterized by the ratio of the rate constants of their addition to a given macroradical: $r_1 = \frac{k_{11}}{k_{12}}$, and $r_2 = \frac{k_{22}}{k_{21}}$. These ratios are referred to as copolymerization constants (r value is always between zero and unity). For instance, for styrene and maleic acid anhydride the copolymerization constants are 0.04 and 0.01, respectively. Sometimes, the same approach is applied to define constants of binary ionic copolymerization.

1. Complete equations of polymerization reactions below and draw structures of compounds $X_1 - X_7$. Give both detailed and short formulas of all copolymers. In short formulas represent styrene units as St, ethylene oxide units as EO, vinyl alcohol units as VA, and maleic anhydride units as MA. Use abbreviations from the above table when necessary.

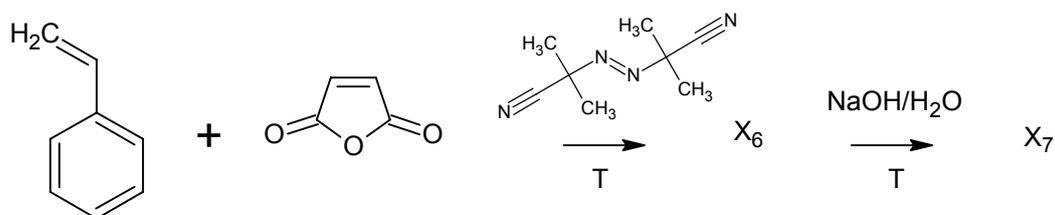
a)



b)



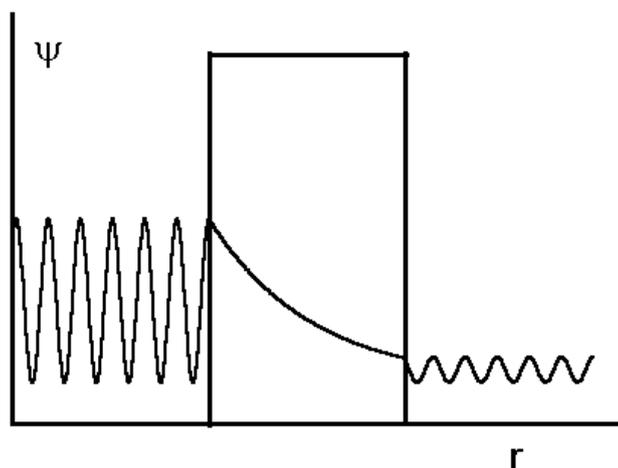
c)



2. Calculate the average length of a chain of units A in the polymer obtained by radical copolymerization of equimolar mixture of two monomers of the same reactivity.

Problem 28. TUNNELING IN CHEMISTRY

Tunneling through energy barriers is a purely quantum-mechanical effect. It is explained by the fact that wave functions can differ from zero even in the classically forbidden areas where energy of a particle is less than an energy barrier:



Inversion of ammonia is a widely known example of tunneling:



In this process the molecule of ammonia is turned out like an umbrella against a strong wind. The tunneling frequency is 24 GHz, and the energy barrier separating two states is 25 kJ/mol.

1. Draw the reaction energy profile (plot of energy vs. reaction coordinate) for the inversion of ammonia. What is the reaction coordinate? What coordinate corresponds to the maximum of energy?
2. In which region of the electromagnetic spectrum can the tunneling of ammonia be observed?
3. Find the energy difference corresponding to the tunneling frequency. What is the ratio of this energy to the barrier height?
4. How would the tunneling frequency change if we substitute some hydrogen atoms by deuterium ones? Explain.

The 39th International Chemistry Olympiad

Chemistry: art, science and fun



PREPARATORY PROBLEMS (Experimental)

July 15-24, 2007

Moscow, Russia

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RULES TO BE FOLLOWED IN LABORATORIES

As mentioned in the Preface, we pay great attention to safety of experimental work. Below you will find a list of rules to be followed during laboratory exam at IChO-2007. We also hope you will take this information into account while preparing for the Olympiad.

- Students have to bring their own laboratory coats.
- Prior to the exam, students will be given Safety instructions in their mother tongue. Each student must carefully read the text and then sign.
- When students enter the lab they must familiarize themselves with the locations of emergency exits, safety shower, fire blanket and eye wash.
- Laboratory coats, eye protections and closed shoes must be worn while staying in the laboratory.
- Coats and bags are forbidden in the laboratory. Those have to be deposited in the cloakroom.
- Eating, drinking or smoking in the laboratory or tasting chemicals are strictly forbidden.
- Pipetting by mouth is strictly forbidden.
- Organizers do their best to avoid harmful chemicals at the exam. All potentially dangerous materials (if any) will be labeled by international symbols. Each student is responsible for recognizing these symbols and knowing their meaning.
- Do not dispose of chemicals down the sink. Follow all disposal instructions provided by Organizers.
- Do not hesitate to ask your lab instructor if you have got any questions regarding safety issues.

Nobody can create rules that will cover all situations, which may happen in reality. We do rely on your common sense and responsibility.

Good luck during preparations and at the exam!

LIST of R- and S-PHRASES

for the reagents used in Experimental problems

R-PHRASES

- R5: Heating may cause an explosion
- R8: Contact with combustible material may cause fire
- R9: Explosive when mixed with combustible material
- R10: Flammable
- R11: Highly flammable
- R20: Harmful by inhalation
- R22: Harmful if swallowed
- R23: Toxic by inhalation
- R25: Toxic if swallowed
- R34: Causes burns
- R35: Causes severe burns
- R36: Irritating to eyes
- R37: Irritating to respiratory system
- R40: Limited evidence of a carcinogenic effect
- R43: May cause sensitization by skin contact
- R50: Very toxic to aquatic organisms
- R61: May cause harm to the unborn child
- R20/21/22: Harmful by inhalation, in contact with skin and if swallowed
- R23/24/25: Toxic by inhalation, in contact with skin and if swallowed
- R36/38: Irritating to eyes and skin
- R36/37/38: Irritating to eyes, respiratory system and skin
- R50/53: Very toxic to aquatic organisms, may cause long-term adverse effects in the aquatic environment

S-PHRASES

- S2: Keep out of the reach of children
- S7: Keep container tightly closed
- S16: Keep away from sources of ignition - No smoking
- S17: Keep away from combustible material
- S22: Do not breathe dust

S23: Do not breathe gas/fumes/vapor/spray (*appropriate wording to be specified by the manufacturer*)

S24: Avoid contact with skin

S26: In case of contact with eyes, rinse immediately with plenty of water and seek medical advice

S28: After contact with skin, wash immediately with plenty of ... (*to be specified by the manufacturer*)

S30: Never add water to this product

S35: This material and its container must be disposed of in a safe way

S36: Wear suitable protective clothing

S37: Wear suitable gloves

S38: In case of insufficient ventilation wear suitable respiratory equipment

S45: In case of accident or if you feel unwell seek medical advice immediately (show the label where possible)

S60: This material and its container must be disposed of as hazardous waste

S61: Avoid release to the environment. Refer to special instructions/safety data sheet

S1/2: Keep locked up and out of the reach of children

S36/37: Wear suitable protective clothing and gloves

S36/37/39: Wear suitable protective clothing, gloves and eye/face protection

S37/39: Wear suitable gloves and eye/face protection

Problem 29. TITRIMETRIC DETERMINATION OF FE IN DIFFERENT OXIDATION STATES

Some methods of iron determination in the oxidation states +2 and +3 are discussed in Problem 12. You are invited to test one more approach to solving that problem in practice.

Reagents and solutions required

KIO₃ (R9, R22, R36/37/38, S35), reagent grade, solid

Ascorbic acid, solid

KI (R36/38, R42-43, R61; S26, S36/37/39, S45), 5% aqueous solution

HCl (R34, R37, S26, S36, S45), conc. and 2 M

HNO₃ (R8, R35, S1/2, S23, S26, S36, S45), conc.

Sulfosalicylic acid, 25% aqueous solution

NH₃ (R10, R23, R34, R50, S1/2, S16, S36/37/39, S45, S61), 10% aqueous solution

EDTA (R36, S26), standard solution, about 0.05 M (the exact value will be given)

1. Preparation of a primary standard solution of KIO₃

1.1. Calculate with the accuracy of 0.0001 g the weight of KIO₃ necessary for the preparation of 200.0 mL of 0.01000 M KIO₃ solution.

1.2. Using analytical balance weigh out accurately a portion of KIO₃. The weight of the portion may differ from the calculated one no more than by 0.05 g and it should be measured with a 0.0001 g accuracy.

1.3. Transfer the portion into 200.0 mL volumetric flask, dissolve it in water, dilute to the mark and mix.

1.4. Calculate the exact concentration of the solution prepared in mol/L.

2. Preparation of the titrant solution – ascorbic acid

2.1. Calculate with the accuracy of 0.01 g the weight of ascorbic acid necessary for preparation of 200 mL of 0.1 M solution.

2.2. Using technical balance weigh out a portion of ascorbic acid. Its weight may differ from the calculated one no more than by 0.05 g.

2.3. Dissolve the portion in ~200 mL of water, mix well, transfer the solution into a vessel and close it tightly with a stopper.

3. Standardization of the ascorbic acid solution

3.1. Fill in a burette with the ascorbic acid solution.

3.2. With a pipette transfer 10.00 mL of standard KIO_3 solution into a 100 mL Erlenmeyer flask, add 20 mL of 5% KI solution and 5 mL of 2 M HCl.

3.3. Titrate the mixture with the ascorbic acid solution until the iodine color disappears.

Note. When titrating iodine with solutions of reducing agents, starch is usually added as an indicator. Here it is not recommended to do so because the reaction rate decreases significantly in presence of starch.

3.4. Repeat the titration until three titrant volumes differ no more than by 0.10 mL.

3.5. Calculate the average titrant volume.

3.6. Calculate the ascorbic acid concentration in the solution in mol/L.

Questions

1. Write down the balanced equations of all the reactions taking place during standardization of ascorbic acid solution. Ascorbic acid $\text{C}_6\text{H}_8\text{O}_6$ is being oxidized to dehydroascorbic acid $\text{C}_6\text{H}_6\text{O}_6$.

2. KIO_3 in presence of excess of KI can be used as a primary standard for HCl standardization as well. The method is similar to that described above with the exception that no HCl is added to the titrated solution in this case. Which compound(s) can be used as an indicator(s) for that titration:

- starch
- sulfosalicylic acid
- methyl orange
- methyl orange + $\text{Na}_2\text{S}_2\text{O}_3$ (in excess)

4. Determination of Fe(III) by ascorbimetric titration

4.1. From your instructor obtain a sample solution containing Fe(II) and Fe(III) (in 100.0-mL volumetric flask). Dilute the solution to the mark with water and mix.

4.2. Fill in the burette with the standardized ascorbic acid solution.

4.3. With a pipette place 10.00 mL of the sample solution into a 100 mL Erlenmeyer flask, add 40 mL of water and heat nearly to boiling.

4.4. Into the hot solution add 4-5 drops of 25% sulfosalicylic acid solution as an indicator.

4.5. Titrate the solution with the ascorbic acid solution until the violet color disappears. During the titration and especially near the end point the solution must be hot. You may need to heat it additionally, if necessary. Near the end point the ascorbic acid solution should be added slowly.

4.6. Repeat the titrations until three titrant volumes differ no more than by 0.10 mL.

4.7. Calculate the average titrant volume.

4.8. Calculate the weight of Fe(III) in the sample solution given to you.

Note. Ascorbic acid, especially in aqueous solutions, is instable and oxidizes with oxygen from the air. Therefore the standardization of ascorbic acid solution and ascorbimetric determination of Fe(III) must be carried out during one workday.

Questions

1. Write down the balanced equations of all the reactions taking place during Fe(III) determination. Ascorbic acid $C_6H_8O_6$ is being oxidized to dehydroascorbic acid $C_6H_6O_6$.

2. In what media does ascorbic acid exhibit its reducing properties most markedly?

- in acidic
- in neutral
- in alkaline
- reducing properties of ascorbic acid do not depend on the pH

5. Determination of total iron by complexometric titration

5.1. Fill in the burette with an EDTA standard solution.

5.2. With a pipette transfer 10.00 mL of the sample solution into a 100 mL Erlenmeyer flask. Add 5 mL of conc. HCl and 2 mL of conc. HNO_3 to oxidize Fe(II) present in the sample to Fe(III). Cover the flask with a watch glass, heat until boiling and continue heating for 3-5 min avoiding splashing.

5.3. Cool down the solution and neutralize it carefully adding 10% NH_3 dropwise until color changes from lemon yellow to yellowish brown and slight turbidity persists.

5.4. Add 1-2 drops of 2 M HCl to dissolve the precipitate, then 0.5 mL of 2 M HCl more, dilute up to 50 mL with distilled water and heat nearly to boiling.

5.5. Into the hot solution add 4-5 drops of 25% sulfosalicylic acid solution as an indicator.

5.6. Titrate the solution until color changes from violet to clear yellow. During the titration and especially near the end point the solution must be hot. You may need to heat it additionally, if necessary. Near the end point the EDTA solution should be added slowly.

5.7. Repeat the titrations until three titrant volumes differ no more than by 0.10 mL.

5.8. Calculate the average titrant volume.

5.9. Calculate the total weight of iron in the sample solution given to you.

5.10. Calculate the weight of Fe(II) as a difference between the results obtained in 5.9 and 4.8.

Questions

1. Write down the balanced equations of all the reactions taking place during total Fe determination.

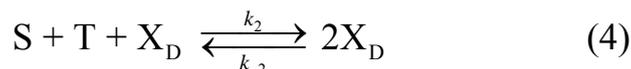
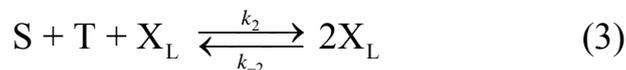
2. One of the crucial items in the Fe(III) determination by complexometric titration is strict maintenance of solution acidity. What are the reasons for that?

- If the acidity is too low, $\text{Fe}(\text{OH})_3$ precipitates
- If the acidity is too high, complex of Fe(III) with sulfosalicylic acid does not form
- If the acidity is too high, complex of Fe(III) with EDTA acid does not form
- If the acidity is too low and/or too high, the titrant decomposes

Problem 30. ASYMMETRIC AUTOCATALYSIS – THE NUMERICAL EXPERIMENT

Nature exhibits a curious asymmetry between the left and the right, which is generally called 'chiral asymmetry'. Indeed, living organisms contain mostly L-amino acids and D-carbohydrates. One of the possible explanations of this phenomenon is based on the idea of autocatalysis. Chiral (asymmetric) autocatalysis is a reaction in which every chiral product serves as the catalyst of its own formation. In such reactions small initial excess of one of the enantiomers can increase exponentially in time.

Consider the kinetic scheme explaining this phenomenon. Two Enantiomers, X_L and X_D , are reversibly formed from achiral reagents T and S:



Enantiomers react with each other giving the product P. The reactions take place in an open system, where constant concentrations of reagents S and T are maintained.

The system of rate equations can be solved numerically using any of the mathematical packages, for example Mathematica, MathCad, etc. Alternatively, you may use the program KINET posted on the official website www.icho39.chem.msu.ru. Let us assume the following values of rate constants (in arbitrary units): $k_1 = 0.5$, $k_{-1} = 0.1$, $k_2 = 0.5$, $k_{-2} = 0.2$, $k_3 = 0.5$.

Procedure

For numerical solution of the systems of differential equations mathematical packages use different commands. In Mathematica it is done by the function NDSolve. The arguments are the list of equations, initial conditions and a time interval. For example, the system of equations

$$\begin{aligned} a'(t) &= -a(t)p(t) \\ p'(t) &= a(t)p(t) - 2 \cdot p(t) \end{aligned}$$

with the initial conditions $a(0) = 2$, $p(0) = 0.5$ in a time interval from $t = 0$ to $t = 10$ is solved numerically by the command:

```
sol=NDSolve [{a' [t]==-a [t]*p [t], p' [t]==a [t]*p [t]-2*p [t], a [0]==2, p [0]==0.5},
  {a, p}, {t, 0,10}]
```

The obtained solution is presented on the graph by the command Plot:

Plot [Evaluate [{a [t], p [t]}/.sol, {t, 0,10}], PlotRange-> All]

Questions

1. Compare equations 1 and 2 or 3 and 4 in the Scheme above. Why are the rate constants identical for enantiomers X_L and X_D ?

2. The control parameter for this problem is the product of concentrations of reagents. Solve the system of kinetic equations numerically and draw on one graph the kinetic curves for X_L and X_D using the initial conditions: $[X_L]_0 = 0$, $[X_D]_0 = 0.01$. Consider two opposite cases: $[S] [T]$ is small, $[S] [T]$ is large. By varying the parameter $[S] [T]$ determine its “break” value at which the shape of kinetic curve(s) changes drastically.

3. At fixed value $[S] [T] = 5$ study the influence of initial chiral asymmetry on kinetic curves. Consider two cases: $[X_D]_0 = 0.001$, $[X_D]_0 = 0.1$.

Let us determine which elementary reactions are essential for chiral asymmetry amplification.

4. Consider the role of reversibility. For this purpose, given the same initial concentrations compare kinetic curves for two mechanisms: with reversible ($k_{-1} \neq 0$; $k_{-2} \neq 0$) and with irreversible formation of the enantiomers ($k_{-1} = k_{-2} = 0$).

5. Consider the simplified scheme in which the first two reactions are absent. Whether or not amplification of chiral asymmetry is possible in such system?

6. Compare the open and closed systems. You have already treated the open system. In the closed system the reagents S and T are no more introduced to a reaction vessel during reaction, therefore they should be included in the system of kinetic equations. Whether or not amplification of chiral asymmetry is possible in a closed system?

Draw the conclusions. What conditions are necessary for amplification of chiral asymmetry to be observed? What elementary stages appear to be essential for it?

Problem 31. OSCILLATING REACTIONS

Introduction

In 1921 W. Bray published an article describing the oscillating reaction of oxidation of hydrogen peroxide with potassium iodate. However thorough investigation of oscillating reaction mechanisms has begun only in 1951, when B.P. Belousov discovered oscillations of concentrations of reduced and oxidized forms of cerium catalyzing oxidation of citric acid by bromate-ion. Later it was shown that oscillating reactions are possible in other redox systems. A.M. Zhabotinsky investigated the oxidation of malonic acid by bromate-ion in the presence of manganese ions. This reaction mechanism is very sophisticated and includes dozens of intermediate compounds.

We will investigate an oscillating reaction taking place in the malonic acid-iodate ion system in the presence of manganese salt and hydrogen peroxide.

Reagents and equipment

- 1) 40 % H_2O_2 (R5, R8, R20, R22, R35; S1/2, S17, S26, S28, S36/37/39, S45)
- 2) KIO_3 (R9, R22, R36/37/38, S35).
- 3) conc. H_2SO_4 (R23/24/25, R35, R36/37/38, R49, S23, S30, S36/37/39, S45)
- 3) $\text{C}_3\text{H}_4\text{O}_4$, malonic acid (R20/21/22, S26, S36/37/39)
- 4) $\text{MnSO}_4 \cdot 5\text{H}_2\text{O}$ (R20/21/22, R36/37/38, R40, S26, S36)
- 5) starch
- 6) KI, solution (R36/38, R42-43, R61; S26, S36/37/39, S45)
- 7) AgNO_3 , solution (R34, R50/53, S1/2, S26, S45, S60, S61)
- 8) analytical balance
- 9) weighing dishes
- 10) flat-bottom flasks or beakers (250-500 ml), 4 items
- 11) stop-watch

Procedure

Prepare three solutions (may be prepared in advance):

- 1) solution of 80 ml 40 % H_2O_2 in 120 ml of water,
- 2) solution of 8.7 g KIO_3 and 0.9 ml conc. H_2SO_4 in 190 ml of water,
- 3) solution of 3 g $\text{C}_3\text{H}_4\text{O}_4$, 2.4 g $\text{MnSO}_4 \cdot 5\text{H}_2\text{O}$ and 0.06 g starch in 195 ml of water.

Mix the solutions in the same vessel and observe the oscillating process. Evaluate the oscillation period and its change in time.

Split the mixture into two parts and place them into beakers.

To one of the parts add AgNO_3 solution (first – several drops, then ~3 ml). Observe changes of the oscillation period. Note the color of the solution upon completion of the oscillation reaction.

To the other part add KI solution (several drops). Observe changes of the oscillation period.

Questions

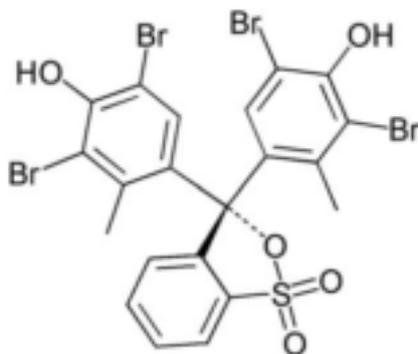
1. Oxidation of malonic acid by potassium iodate is an autocatalytic process. Write down the net equation of the reaction. Which product is the catalyst of the oscillating process? Explain the effect of silver nitrate.
2. B.P. Belousov used bromate-ion as an oxidizing agent. Suggest what would happen if we substitute iodate-ion by bromate-ion in the reaction with malonic acid. What role does hydrogen peroxide play in the oxidation of malonic acid with iodate-ion?
3. It is well known, that one of the stages of the oscillating process is formation of iodomalonic acid with its subsequent decomposition. How can we explain the fact that potassium iodide inhibits the reaction?
4. B.P. Belousov used the $\text{Ce}^{4+}/\text{Ce}^{3+}$ redox couple to study oscillating reactions. Is it possible to use the following transient metal redox couples as a catalyst: $\text{Co}^{3+}/\text{Co}^{2+}$, $\text{Fe}^{3+}/\text{Fe}^{2+}$, $\text{Tl}^{3+}/\text{Tl}^{1+}$?

$$E^\circ(\text{Co}^{3+}/\text{Co}^{2+}) = 1.81 \text{ V}, E^\circ(\text{Ce}^{4+}/\text{Ce}^{3+}) = 1.61 \text{ V},$$

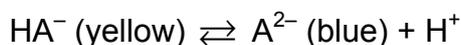
$$E^\circ(\text{Mn}^{3+}/\text{Mn}^{2+}) = 1.51 \text{ V}, E^\circ(\text{Fe}^{3+}/\text{Fe}^{2+}) = 0.77 \text{ V?}$$

Problem 32. DETERMINATION OF THE ACIDITY CONSTANT OF BROMOCRESOL BLUE (3',3'',5',5''-TETRABROMO-M-CRESOLSULFONEPHTHALEIN, BCB)

Bromocresol blue (BCB)



is an organic dye, an acid-base indicator, a weak diprotic acid (H_2A). In aqueous solutions in the pH range of 3-6 BCB changes its color from yellow to blue due to dissociation of the second proton:



On the base of the absorbance of BCB solution measured as a function of the pH one can calculate the second acidity constant of BCB, $\text{p}K_{a2}$.

Reagents and solutions required

Bromocresol blue, 0.25% solution in 50% aqueous ethanol (R11, S2, S7, S16).

Mixture of acids for preparation of buffer solutions: an aqueous solution containing H_3PO_4 , (R34, S1/2, S26, S45), CH_3COOH (R10, R35, S1/2, S23, S26, S45) and H_3BO_3 , (S22, S26, S36/37, S38, S45), 0.04 M each.

NaOH (R35, S1/2, S26, S37/39, S45), 0.2 M and 2 M solutions.

HCl (R34, R37, S26, S36, S45), 2 M solution.

1. Choice of the wavelength for the K_{a2} determination

1.1. Into each of two 50.0 mL volumetric flasks place 1.00 mL of the BCB solution and 10.00 mL of the mixture of acids (see reagent list). Then add 1.00 mL of 0.2 M NaOH

into the first and 6.00 mL of 2 M NaOH into the second flask. Dilute the solutions to the mark with water and mix.

1.2. Measure the pH of the solutions prepared. The first one must have the pH in the range of 2-3, the second – within 7-8. Under such conditions all BCB is in the form of HA^- or A^{2-} respectively. If either of the pH is different from the required, adjust it by adding few drops of 2 M HCl or 2 M NaOH.

1.3. Measure the absorption spectra of the solutions in the range of 400-700 nm; 5-10 data points would be sufficient.

1.4. Choose the wavelength at which the absorbances of the solutions differ most greatly. Usually that wavelength corresponds to the maximum of absorbance of one of the species or close to it. Further carry out all the measurements at that wavelength.

2. Preparation of series of BCB solutions, measuring their absorbance and the pH

2.1. Into each of twelve 50-mL volumetric flasks place 1.00 mL of BCB solution and 10.00 mL of the mixture of acids. Then add 0.2 M NaOH to each flask in the amount indicated in Table below:

| Flask number | 0,2 M NaOH, mL |
|--------------|----------------|
| 1 | 0.75 |
| 2 | 1.50 |
| 3 | 2.50 |
| 4 | 2.75 |
| 5 | 3.00 |
| 6 | 3.25 |
| 7 | 3.50 |
| 8 | 3.75 |
| 9 | 4.00 |
| 10 | 4.25 |
| 11 | 5.25 |
| 12 | 6.25 |

Dilute the solutions to the mark with water and mix.

Note. It is of essential importance that the concentrations of BCB be strictly the same in all the solutions. When preparing the solutions pay especial attention to that requirement!

2.2. For each solution measure the pH and the absorbance at the chosen wavelength.

2.3. Using the data obtained calculate $\log K_{a2}$ for each of the solutions unless fraction of either of the species involved in the acid-base equilibrium is negligible.

2.4. Calculate the average $\log K_{a2}$ value.

Questions

Denote as:

$[HA^-]$, $[A^{2-}]$, c – equilibrium concentrations of the corresponding BCB forms and its total concentration, respectively;

l – cuvette length;

K_{a2} – acidity constant of HA^- ;

ε_{HA} , ε_A – extinction coefficients of the corresponding forms at the chosen wavelength;

A_{HA} , A_A , A – absorbances of BCB solution containing only HA^- , only A^{2-} and their mixture, respectively.

1. Write down the equations for A_{HA} , A_A and A as functions of $[HA^-]$, $[A^{2-}]$ and c .

2. Express A as a function of A_{HA} , A_A and $[H^+]$.

3. Write down the equation for calculation of K_{a2} from A_{HA} , A_A , A and $[H^+]$.

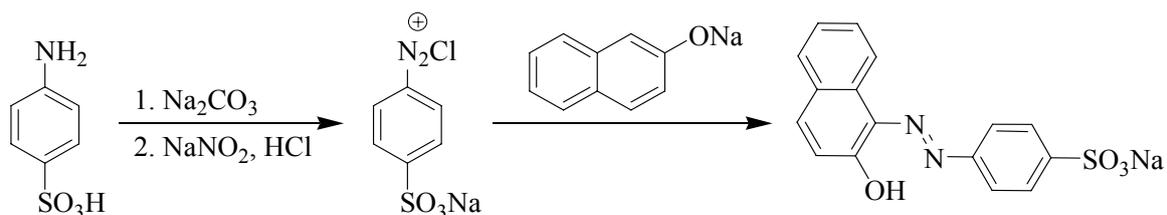
4. Consider the wavelength at which $\varepsilon_{HA} = \varepsilon_A$. It is called the isosbestic point.

a) Is it possible to determine K_a of a dye by measuring the absorbance at the isosbestic point?

b) What analytical information can be obtained from such measurement?

Problem 33. ACID ORANGE 7

A very popular azo-dye known under dozens of trade names and widely used in textile, leather, food, cosmetics, as well as other industries, Acid Orange 7 (Acid Orange II, Persian Orange, listed in the Color Index as No. 15510) can be readily obtained by azo-coupling of diazotized sulphanylic acid with 2-naphtholate

**Materials and hardware**

Sulfanylic acid (R36/37/38, R43, S24, S37)

2-Naphthol (R36/37/38, S26, S37)

Sodium carbonate (R36, S2, S22, S26)

Sodium nitrite (R8, R25, R36/37/38, R50, S26, S36, S45, S61)

Sodium hydroxide (R35, S1/2, S26, S37/39, S45)

Hydrochloric acid, conc. (R34, R37, S26, S36, S45)

Ice

Glass beakers (150, 200, 500 ml), thermometer, spatulas, magnetic stirrer and heating plate, vacuum filtration apparatus, desiccator.

The diazotization

Sulfanylic acid (8.66 g, 0.05 mol) is dissolved in the solution of 3 g of sodium carbonate in 50 ml water in a 150 ml glass beaker placed on a magnetic stirrer. 15 ml of concentrated HCl are added to this solution at vigorous stirring. After cooling to room temperature, the beaker is immersed in an ice bath (a couple of ice chunks can be added to the mixture to ensure good cooling) and the mixture is further cooled to 0 °C. A solution of NaNO_2 (3.45 g, 0.05 mol) in 20 ml of water is added dropwise (**warning!** this

operation should be done in a hood because of evolution of nitrogen oxides). The rate of addition should be controlled to keep the temperature near 0 °C as accurately as possible (**warning!** even a 2-3° increase leads to side-reactions which may lead to the formation of phenols giving unwanted azo-dyes which dramatically worsen the purity of color of the target dye). During the addition white precipitate of diazonium salt (diazotized sulfanilate is a betaine, an inner salt with zero net charge, therefore it is not well soluble in water) may sometimes form. The results of diazocoupling do not depend on whether the diazonium salt is in solution or suspension.

After the addition of all nitrite solution, stirring is continued for 10-15 min (**warning!** temperature should be carefully controlled!). The diazonium salt solution (or suspension) should be used immediately after preparation.

The azocoupling

2-Naphthol (7.21 g, 0.05 mol) is dissolved in 40 ml of 5% NaOH solution. This solution is mixed with solution of 12.5 g Na₂CO₃ in 100 ml water in a 500 ml beaker. The resulting solution should be transparent, if any precipitate or suspension persists, it should be filtered off. The solution of naphtholate is cooled to 0 °C by ice (an ice bath + a few ice chunks inside). The diazonium salt solution is slowly poured to naphtholate solution under vigorous stirring by a spatula or a glass rod. Attention should be paid to keep the temperature below 8 °C throughout the addition. Afterwards, the mixture is left for an hour, preferably on a magnetic stirrer. The dye partially precipitates as golden plates. After an hour, the solution is heated to completely dissolve the precipitate, filtered hot (*note*: this filtration can be omitted if a hot filtration funnel is not available), and saturated by 50 g of sodium chloride (50 g) while hot (it is necessary to keep temperature above 50° during saturation, so the beaker should be placed on a heating plate). Dye precipitate formed by salting-out is filtered off by vacuum filtration from hot solution (*note*: if the temperature of solution being filtered drops below 50°, sodium chloride partially co-precipitates with the dye). The dye is dried in a desiccator over CaCl₂. Orange solid, yield 25 g.

The quality of dye can be controlled by the UV/Vis spectroscopy. In aqueous solution λ_{\max} 487 nm (log ϵ 4.87).

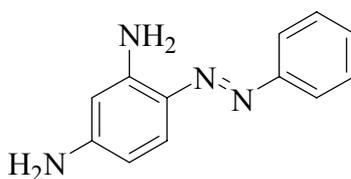
Questions

1. Under the name *tropaeolin 000* the dye is used as an acid-base indicator in aqueous solutions. Guess in which region of pH this dye changes its color:

strongly acidic (pH<2); acidic (pH 2-6.5); neutral (pH 6.5-7.5) mildly alkaline (pH 7.5-9); strongly alkaline (pH 9-14).

2. Write the reaction equation which accounts for the color change.

3. Write the reaction equation of an azocoupling required to obtain *chrysoidine* dye.



4. Which pH region should be chosen for this azocoupling:

strongly basic, weakly basic, weakly acidic, strongly acidic?

PROBLEM 34. DETERMINATION OF MOLECULAR WEIGHT OF A PROTEIN USING GEL FILTRATION

Gel filtration is a simple and reliable chromatographic method for separating molecules according to their size. Within a fractionation range chosen, molecules are eluted in a decreasing order of their size. Versatility of the method makes it applicable for purification and characterization of biological substances of all classes, including macromolecules not readily fractionated by other techniques.

Some gel forming organic polymers with a 3D network structure (usually referred to as gel filtration media, GFM) possess properties of molecular sieves and can separate molecules according to their size and shape. A chromatography column should be filled with swollen gel and equilibrated with corresponding buffer solution. The separation mechanism is non-adsorptive and independent of the eluent system used, thus being

fairly gentle. Liquid inside porous gel beads of GFM is the stationary phase, whereas eluent solution outside the beads is the mobile one.

In a column, all sample molecules can be present in the liquid between the beads. The total volume of such “outside” liquid is referred to as *the void volume* in gel filtration and is equal to about 30% of the column volume. Sample molecules are partitioned between the eluent (the mobile phase) and the accessible part of bead pores (the stationary phase). This partitioning acts to establish a *dynamic equilibrium* of sample molecules between the mobile and stationary phases and is driven exclusively by diffusion. The mobile phase transports the sample molecules down the column. The molecules present in the pores are “stationary” and not subjected to transportation. Migration rate of a sample zone depends on the fraction of sample molecules present in the mobile phase. Separation of individual macromolecules can only be achieved in the case of their partial access to the pores of the GFM. *Applicable sample volume* is restricted to 0.5-5% of that of the column, since no concentration effect is active in gel filtration. *Flow rate* is kept low to avoid peak broadening due to incomplete mass transfer, whereas columns used are long to allow optimum resolution.

Materials

Blue dextran (molecular weight, MW=2 MDa), 4 mg

Proteins:

Ovalbumin (MW=43 kDa), 1.5 mg

Cytochrome C (MW=13 kDa), 0.4 mg

Bovine serum albumin (BSA) (MW=67 kDa), 2.2 mg

Chymotrypsinogen (MW=25 kDa), 1 mg

Hemoglobin (MW=64.5 kDa), 1.5 mg

0.1 M HCl (R34, R37, S26, S36, S45) 230 mL, KCl 22.35 g

Buffer: Tris (2-Amino-2-(hydroxymethyl)propane-1,3-diol; R36/37/38, S26, S36) 6.05 g

GFM: Toyopearl HW-50 (or HW-55), fine, 70 mL.

If the mentioned above proteins are partially inaccessible, those missing can be substituted by proteins with close MW, but not proteases. Toyopearl may be also replaced by a GFM with similar properties.

Apparatus

70 mL chromatography column; packing reservoir; stand; peristaltic pump; UV-cord connected to plotter; Eppendorf centrifuge; analytical balances; water-jet pump; one 1000 mL measuring cylinder; one 250 mL volumetric flask; one big Buchner funnel with glass filter; one 1000 mL Bunsen flask; one 1000 mL round-bottom flask; one 100 μL micropipette with tips; one 1000 μL micropipette with tips; one 2 mL syringe connected to 20 cm tubing; four Eppendorf tubes; one 100 mL measuring cylinder; one 200 mL flask; one 100 mL beaker; big steel spatula; small spatula; glass rod; filter paper.

Note: A UV-cord can be substituted by a UV-visible spectrophotometer and measuring test tubes.

Procedure

Step 1. Preparation of buffer solution

To prepare 0.2 M Tris buffer solution, dissolve 6.05 g of Tris in 250 mL of distilled water in the 250 mL volumetric flask. Mix 125 mL of 0.2 M Tris solution and 230 mL of 0.1 M HCl in the 1000 mL measuring cylinder. Add distilled water to 800 mL. Add 22.35 g of KCl to the Tris-HCl solution and stir thoroughly until the salt completely dissolves. Add water to 1000 mL (the final concentration of KCl is 0.3 M).

Step 2: Preparation of a chromatographic column

Packaging the column is one of the most important stages in chromatography, as it determines the separation quality to a great extent. The column should be packed uniformly, and the upper and lower gel surfaces should be strictly horizontal.

1. Equilibrate gel material to room temperature.
2. Gently shake the bottle to make an even slurry.
3. Pour 70 mL of gel slurry into a beaker and dilute with buffer to 100 mL.
4. Stir with a glass rod to make a homogeneous suspension free from aggregates.
5. Add eluent buffer solution to the column to check for leaks, wet the walls of the column and remove air from the bed support. (It is better to fill the column bottom-up)

using the water-jet pump). Drain buffer leaving about 1 cm above the gel surface. For columns with bottom glass porous filter, a filter paper circle with a diameter equal to the inner column diameter should be placed on the glass filter to prevent from gel leakage from the column.

6. Mount the column vertically and attach the addition packing reservoir firmly to the column. It should be twice shorter than the column.

7. Wash the gel with three portions (of about 100-120 mL) of Tris-buffer solution on Buchner funnel with glass filter attached to 1000 mL Bunsen flask using water-jet pump. Try not to dry Toyopearl. After each washing disconnect the water-jet pump when the upper gel surface just starts turning dry. Then add next portion of buffer, stir with big steel spatula to make a homogeneous suspension, and subject to suction.

8. Transfer the gel from the funnel into 1000 mL round-bottom flask, add 50 mL of buffer solution and connect the flask to water-jet pump using a connector. Vacuum degassing should proceed for at least 5 min.

9. Re-suspend and pour the gel slurry into the column in one continuous motion. Pouring down a glass rod held against the wall of the column prevents from air bubbles (Fig.1). Try gel slurry to flow along the column wall.

10. Carefully fill the reservoir to the top with buffer solution, disturbing the gel as little as possible. Connect the reservoir with the peristaltic pump, which should in turn be joined to buffer stock in the 200 mL flask. Turn on the pump and open the column outlet.

11. Buffer solution should be pumped through the column until the gel stops settling. After two bed volumes remove the gel reservoir and insert flow adaptor.

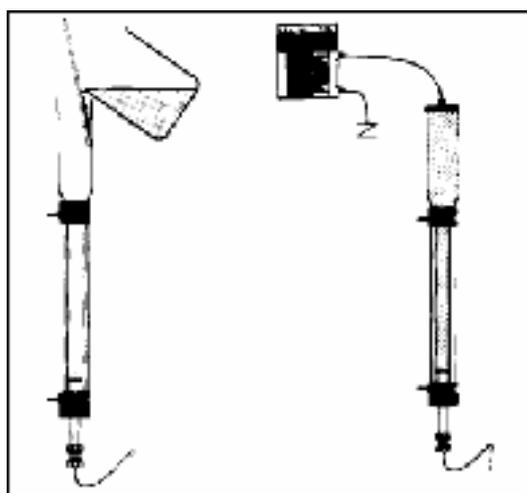


Fig. 1. Packaging the column with GFM.

Step 3: Preparation of solutions

Weigh blue dextran and proteins using balance and small spatula. Prepare solution of Blue dextran by dissolving it in 1 mL of Tris-buffer solution in an Eppendorf tube. Prepare two solutions of standard proteins in Eppendorf tubes. The first solution contains Ovalbumin, Cytochrome C, 0.07 mL of blue dextran solution and 0.93 mL of Tris-buffer solution. The second solution contains Bovine serum albumin, Chymotrypsinogen, 0.07 mL of blue dextran solution and 0.93 mL of Tris-buffer solution. Prepare solution of Hemoglobin (unknown protein) in 1 mL of Tris-buffer solution. Centrifuge two solutions with standard proteins and the solution of unknown protein for 5 min.

Step 4: Application of samples

1. Apply sample solutions carefully, trying not to disturb the gel. To make it easier, filter paper circle could be placed at the top of gel (still take into account possible protein absorption on the paper). Remove flow adaptor, disconnect the peristaltic pump and open the column outlet. Let the buffer soak into the gel (the gel surface should be free of buffer but not dry) and close the column outlet. Add sample solution slowly using pipette with wide tip or 2 mL syringe connected to 20 cm tubing, open the column outlet and allow the solution flow inside the gel. Close the column outlet and add buffer solution (about 1 mL) slowly and carefully (as during the sample application). Open the column outlet and let the buffer soak in the gel. Repeat the procedure. This allows the sample solution flowing deeper inside the gel and prevents from backward diffusion. Close the column outlet and carefully make a buffer layer with height of about 2 cm over the gel.

2. Connect the peristaltic pump to the column inlet and the UV-cord to the column outlet (the tube length should be as short as possible) and start elution.

Step 5: Column chromatography

1. Carry out calibration of the column in two steps:

A. Apply the first solution of standard proteins containing Blue dextran, Ovalbumin and Cytochrome C to the column. Start elution with the rate of about 1-2 mL/min, collecting

the eluate into 100 mL measuring cylinder. The elution process is monitored by following the eluate absorbance at 280 nm, which is registered by the UV-cord. Measure Elution volumes for Blue dextran and proteins using cylinder (record the volumes corresponding to maxima of the eluate absorbance).

Note: in the case of using a spectrophotometer and test-tubes, the procedure should be modified as follows. Collect the eluate in a measuring cylinder up to 25% of the column volume. Then continue collecting the eluate in test-tubes in portions of 1 mL. Determine the eluate absorbance at 280 nm in each test-tube by using a spectrophotometer and record the total volumes corresponding to maxima of the eluate absorbance).

After the three peaks are registered, the column should be washed with the buffer solution until the total elution volume becomes equal to that of the column.

B. Apply the second solution of standard proteins and proceed as described above.

2. Apply the solution of unknown protein. After the peak is registered, stop the peristaltic pump, close column outlet and turn off the UV-cord.

Questions

1. Correlate chromatographic peaks with substances you applied to the column. Complete the table:

| Standard solution number | Number of peak (in the order of appearance) | | |
|--------------------------|---|---|---|
| | 1 | 2 | 3 |
| 1 | | | |
| 2 | | | |

2. What is the void volume of your column? Explain.

3. Calculate the volume of the chromatographic column.

4. Calculate the availability coefficient K_{av} for all proteins using formula

$$K_{av} = \frac{V_r - V_0}{V_c - V_0}$$

V_r is elution volume for sample molecule, V_0 is the void volume, V_c is the column volume.

5. Plot the calibration curve as the dependence of K_{av} on $\log(\text{MW})$ using the data obtained for four standard proteins.

6. Determine MW for the unknown protein.

7. Another important characteristic of a column is *the exclusion limit*, M_r , which is defined as the molecular mass of the smallest molecule excluded from the pores. Calculate this parameter by finding the intercept of the extrapolated linear part of the calibration curve with the $\log(\text{MW})$ axis.

8. Estimate the elution volume for low molecular weight substances if applied to the column under consideration. Provide an explanation.